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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶ : C12N 15/55, 9/16, 5/12, 1/19, 1/21, C07K 16/40, 16/42, G01N 33/566, C12Q 1/68, A61K 38/17	A2	(11) International Publication Number: WO 99/42596 (43) International Publication Date: 26 August 1999 (26.08.99)
(21) International Application Number: PCT/US99/03879 (22) International Filing Date: 23 February 1999 (23.02.99) (30) Priority Data: 60/075,508 23 February 1998 (23.02.98) US (71) Applicant: ICOS CORPORATION [US/US]; 22021 20th Avenue, S.E., Bothell, WA 98021 (US). (72) Inventor: LOUGHNEY, Kate; 7301 34th Avenue, N.E., Seattle, WA 98115 (US). (74) Agent: WILLIAMS, Joseph, A., Jr.; Marshall, O'Toole, Gerstein, Murray & Borun, 6300 Sears Tower, 233 South Wacker Drive, Chicago, IL 60606-6402 (US).		(81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, UZ, VN, YU, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG). Published <i>Without international search report and to be republished upon receipt of that report.</i>
(54) Title: PHOSPHODIESTERASE 10 (57) Abstract The present invention provides novel human PDE10 polypeptides, polynucleotides encoding the polypeptides, expression constructs comprising the polynucleotides, host cells transformed with the expression constructs; methods for producing PDE10 polypeptides; antisense polynucleotides; and antibodies specifically immunoreactive with the PDE10 polypeptides. The invention further provides methods to identify binding partners of PDE10, and more particularly, binding partners that modulate PDE10 enzyme activity.		

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PHOSPHODIESTERASE 10

This application claims priority of U.S. Provisional Application No. 60/075,508, filed February 23, 1998.

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FIELD OF THE INVENTION

The present invention relates generally to a novel phosphodiesterase (PDE) designated PDE10. Depending on nomenclature used, PDE10 is also referred to as PDE9.

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BACKGROUND OF THE INVENTION

Phosphodiesterases (PDEs) hydrolyze 3', 5' cyclic nucleotides to their respective nucleoside 5' monophosphates. The cyclic nucleotides cAMP and cGMP are synthesized by adenylyl and guanylyl cyclases, respectively, and serve as second messengers in a number of cellular signaling pathways. The duration and strength of the second messenger signal is a function of the rate of synthesis and the rate of hydrolysis of the cyclic nucleotide.

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Multiple families of PDEs have been identified. The nomenclature system includes first a number that indicates the PDE family. To date, nine families (PDE1-9) are known which are classified by: (i) primary structure; (ii) substrate preference; (iii) response to different modulators; (iv) sensitivity to specific inhibitors; and (v) modes of regulation [Loughney and Ferguson, in Phosphodiesterase Inhibitors, Schudt, *et al.* (Eds.), Academic Press: New York, New York (1996) pp. 1-19]. The number indicating the family is followed by a capital letter, indicating a distinct gene, and the capital letter followed by a second number, indicating a specific splice variant or a specific transcript that utilizes a unique transcription initiation site.

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The amino acid sequences of all mammalian PDEs identified to date include a highly conserved region of approximately 270 amino acids located in the carboxy terminal half of the protein [Charbonneau, *et al.*, *Proc. Natl. Acad. Sci. (USA)* 83:9308-9312 (1986)]. The conserved domain includes the catalytic site for cAMP and/or cGMP hydrolysis and two putative zinc binding sites as well as family specific determinants [Beavo, *Physiol. Rev.* 75:725-748 (1995); Francis, *et al.*, *J. Biol. Chem.*

-2-

269:22477-22480 (1994)]. The amino terminal regions of the various PDEs are highly variable and include other family specific determinants such as: (i) calmodulin binding sites (PDE1); (ii) non-catalytic cyclic GMP binding sites (PDE2, PDE5, PDE6); (iii) membrane targeting sites (PDE4); (iv) hydrophobic membrane association sites (PDE3); and (v) phosphorylation sites for either the calmodulin-dependent kinase II (PDE1), the cAMP-dependent kinase (PDE1, PDE3, PDE4), or the cGMP dependent kinase (PDE5) [Beavo, *Physiol. Rev.* 75:725-748 (1995); Manganiello, *et al.*, *Arch. Biochem. Acta* 322:1-13 (1995); Conti, *et al.*, *Physiol. Rev.* 75:723-748 (1995)].

Members of the PDE1 family are activated by calcium-calmodulin. Three genes have been identified; PDE1A and PDE1B preferentially hydrolyze cGMP while PDE1C has been shown to exhibit a high affinity for both cAMP and cGMP. The PDE2 family is characterized as being specifically stimulated by cGMP [Loughney and Ferguson, *supra*]. Only one gene has been identified, PDE2A, the enzyme product of which is specifically inhibited by erythro-9-(2-hydroxy-3-nonyl)adenine (EHNA). Enzymes in the PDE3 family are specifically inhibited by cGMP. Two genes are known, PDE3A and PDE3B, both having high affinity for both cAMP and cGMP, although the V_{max} for cGMP hydrolysis is low enough that cGMP functions as a competitive inhibitor for cAMP hydrolysis. PDE3 enzymes are specifically inhibited by milrinone and enoximone [Loughney and Ferguson, *supra*]. The PDE4 family effects cAMP hydrolysis and includes four genes, PDE4A, PDE4B, PDE4C, and PDE4D, each having multiple splice variants. Members of this family are specifically inhibited by the anti-depressant drug rolipram. Members of PDE5 family bind cGMP at non-catalytic sites and preferentially hydrolyze cGMP. Only one gene, PDE5A, has been identified. The photoreceptor PDE6 enzymes specifically hydrolyze cGMP [Loughney and Ferguson, *supra*]. Genes include PDE6A and PDE6B (the protein products of which dimerize and bind two copies of a smaller γ inhibitory subunit to form rod PDE), in addition to PDE6C which associates with three smaller proteins to form cone PDE. The PDE7 family effects cAMP hydrolysis but, in contrast to the PDE4 family, is not inhibited by rolipram [Loughney and Ferguson, *supra*]. Only one gene, PDE7A, has been identified. The PDE8 family has been shown to hydrolyze both cAMP and cGMP and is insensitive to inhibitors specific for PDEs 1-5. Depending on nomenclature used, PDE8 is also referred to as PDE10, but is distinct from

-3-

PDE10 described herein. The PDE9 family preferentially hydrolyzes cAMP and is not sensitive to inhibition by rolipram, a PDE4-specific inhibitor, or isobutyl methyl xanthine (IBMX), a non-specific PDE inhibitor. Depending on nomenclature used, PDE9 is also referred to as PDE8, but is distinct from PDE8 mentioned above. To date, two genes have been identified in the PDE9 family.

Specific and non-specific inhibitors of the various PDE protein families have been shown to be effective in treating disorders attributable, in part, to aberrant levels of cAMP or cGMP. For example, the PDE4-specific inhibitor rolipram, mentioned above as an anti-depressant, inhibits lipopolysaccharide-induced expression of $\text{TNF}\alpha$ and has been effective in treating multiple sclerosis in an animal model. Other PDE4-specific inhibitors are being investigated for use as anti-inflammatory therapeutics, and efficacy in attenuating the late asthmatic response to allergen challenge has been demonstrated [Harbinson, *et al.*, *Eur. Respir. J.* 10:1008-1014 (1997)]. Inhibitors specific for the PDE3 family have been approved for treatment of congestive heart failure. PDE5 inhibitors are currently being evaluated for treatment of penile erectile dysfunction [Boolell, *et al.*, *Int. J. Impotence Res.* 8:47-50 (1996)]. Non-specific inhibitors, such as theophylline and pentoxifylline, are currently used in the treatment of respiratory and vascular disorders, respectively.

Given the importance of cAMP and cGMP in intracellular second messenger signaling, there thus exists an ongoing need in the art to identify additional PDE species. Identification of heretofore unknown families of PDEs, and genes and splice variants thereof, will provide additional pharmacological approaches to treating conditions in which cyclic nucleotide pathways are aberrant, as well as conditions in which modulation of intracellular cAMP and/or cGMP levels in certain cell types is desirable. Identification of family-specific and enzyme-specific inhibitors will permit development of therapeutic and prophylactic agents which act on desired cell types expressing PDEs and/or particular metabolic pathways regulated by cyclic nucleotide monophosphate steady-state concentrations.

SUMMARY OF THE INVENTION

In brief, the present invention provides purified and isolated PDE10 polypeptides. Preferred polypeptides comprise the amino acid sequence selected from the

-4-

group consisting of SEQ ID NO: 2, SEQ ID NO: 18, SEQ ID NO: 20 and SEQ ID NO: 22.

The invention also provides polynucleotides encoding polypeptides of the invention. A preferred polynucleotide comprises the sequence set forth in SEQ ID NO:

5 1. Polynucleotides of the invention include polynucleotides encoding a human PDE10 polypeptide selected from the group consisting of: a) the polynucleotide according to SEQ ID NO: 1, 18, 20 or 22; b) a DNA which hybridizes under moderately stringent conditions to the non-coding strand of the polynucleotide of (a); and c) a DNA which would hybridize to the non-coding strand of the polynucleotide of (a) but for the
10 redundancy of the genetic code. Polynucleotides of the invention comprise any one of the polynucleotide set out in SEQ ID NO: 18, SEQ ID NO: 20, and SEQ ID NO: 22, as well as fragments thereof. The invention provide polynucleotides which are DNA molecules. DNA molecules include cDNA, genomic DNA, and wholly or partially chemically synthesized DNA molecule. The invention also provides antisense polynucleotides which
15 specifically hybridizes with the complement of a polynucleotide of the invention.

The invention also provides expression constructs comprising a polynucleotide of the invention, host cells transformed or transfected with an expression construct of the invention, and methods for producing a PDE10 polypeptide comprising the steps of: a) growing the host cell of the invention under conditions appropriate for
20 expression of the PDE10 polypeptide and b) isolating the PDE10 polypeptide from the host cell or the medium of its growth.

The invention further provides antibodies specifically immunoreactive with a polypeptide of the invention. Preferably, the antibody is a monoclonal antibody. The invention also provides hybridomas which produces an antibody of the invention.
25 Anti-idiotypic antibody specifically immunoreactive with the antibody of the invention are also contemplated.

The invention also provides methods to identify a specific binding partner compound of a PDE10 polypeptide comprising the steps of: a) contacting the PDE10 polypeptide with a compound under conditions which permit binding between the
30 compound and the PDE10 polypeptide; b) detecting binding of the compound to the PDE10 polypeptide; and c) identifying the compound as a specific binding partner of the

-5-

PDE10 polypeptide. Preferably, methods of the invention identify specific binding partners that modulate activity of the PDE10 polypeptide. In one aspect, the methods identify compounds that inhibit activity of the PDE10 polypeptide. In another aspect, the methods identify compounds that enhance activity of the PDE10 polypeptide.

5 The invention also provides methods to identify a specific binding partner compound of the PDE10 polynucleotide of the invention comprising the steps of: a) contacting the PDE10 polynucleotide with a compound under conditions which permit binding between the compound and the PDE10 polynucleotide; b) detecting binding of the compound to the PDE10 polynucleotide; and c) identifying the compound as a specific
10 binding partner of the PDE10 polynucleotide. Preferably, the methods identify specific binding partner compounds that modulate expression of a PDE10 polypeptide encoded by the PDE10 polynucleotide. In one aspect, method of the invention identify compounds that inhibit expression of the PDE10 polypeptide. In another aspect, methods of the invention identify compounds that enhance expression of the PDE10 polypeptide.

15 Binding partner compounds identified by methods of the invention are also contemplated, as are compositions comprising the compound. The invention further comprehends use of binding partner compounds of the invention in production of medicaments for the treatment of PDE10-related disorders.

20 DETAILED DESCRIPTION OF THE INVENTION

 The present invention provides polypeptides and underlying polynucleotides for a novel PDE family designated PDE10. The PDE10 family is distinguished from previously known PDE families in that it shows a lower degree of sequence homology than would be expected for a member of a known family of PDEs and
25 it is not sensitive to inhibitors that are known to be specific for previously identified PDE families. Outside of the catalytic region of the protein, PDE10 shows little homology to other known PDEs. Even over the catalytic region, PDE10 amino acid sequence identity is less than 40% when compared with the same region in known PDEs. The invention includes both naturally occurring and non-naturally occurring PDE10 polynucleotides and
30 polypeptide products thereof. Naturally occurring PDE10 products include distinct gene and polypeptide species within the PDE10 family; these species include those which are

-6-

expressed within cells of the same animal as well as corresponding species homologs expressed in cells of other animals. Within each PDE10 species, the invention further provides splice variants encoded by the same polynucleotide but which arise from distinct mRNA transcripts. Non-naturally occurring PDE10 products include variants of the naturally occurring products such as analogs (*i.e.*, wherein one or more amino acids are added, substituted, or deleted) and those PDE10 products which include covalent modifications (*i.e.*, fusion proteins, glycosylation variants, and the like).

The present invention provides novel purified and isolated polynucleotides (e.g., DNA sequences and RNA transcripts, both sense and complementary antisense strands, including splice variants thereof) encoding human PDE10s. DNA sequences of the invention include genomic and cDNA sequences as well as wholly or partially chemically synthesized DNA sequences. Genomic DNA of the invention comprises the protein coding region for a polypeptide of the invention and includes allelic variants of the preferred polynucleotide of the invention. Genomic DNA of the invention is distinguishable from genomic DNAs encoding polypeptides other than PDE10 in that it includes the PDE10 coding region as defined by PDE10 cDNA of the invention. The invention therefore provides structural, physical, and functional characterization for genomic PDE10 DNA. Allelic variants are known in the art to be modified forms of a wild type gene sequence, the modification resulting from recombination during chromosomal segregation or exposure to conditions which give rise to genetic mutation. Allelic variants, like wild type genes, are inherently naturally occurring sequences (as opposed to non-naturally occurring variants which arise from *in vitro* manipulation). "Synthesized," as used herein and is understood in the art, refers to purely chemical, as opposed to enzymatic, methods for producing polynucleotides. "Wholly" synthesized DNA sequences are therefore produced entirely by chemical means, and "partially" synthesized DNAs embrace those wherein only portions of the resulting DNA were produced by chemical means. A preferred DNA sequence encoding a human PDE10 polypeptide is set out in SEQ ID NO: 1. The worker of skill in the art will readily appreciate that the preferred DNA of the invention comprises a double stranded molecule, for example the molecule having the sequence set forth in SEQ ID NO: 1 along with the complementary molecule (the "non-coding strand" or "complement") having a sequence

-7-

deducible from the sequence of SEQ ID NO: 1 according to Watson-Crick base pairing rules for DNA. Also preferred are polynucleotides encoding the PDE10 polypeptide of SEQ ID NO: 2.

5 The disclosure of a full length polynucleotide encoding a PDE10 polypeptide makes readily available to the worker of ordinary skill in the art every possible fragment of the full length polynucleotide. The invention therefore provides fragments of PDE10-encoding polynucleotides of the invention comprising at least 10 to 20, and preferably at least 15, nucleotides, however, the invention comprehends fragments of various lengths. Preferably, fragment polynucleotides of the invention comprise sequences
10 unique to the PDE10-encoding polynucleotide sequence, and therefore hybridize under stringent or preferably moderate conditions only (*i.e.*, "specifically") to polynucleotides encoding PDE10, or PDE10 polynucleotide fragments containing the unique sequence. Polynucleotide fragments of genomic sequences of the invention comprise not only sequences unique to the coding region, but also include fragments of the full length
15 sequence derived from introns, regulatory regions, and/or other non-translated sequences. Sequences unique to polynucleotides of the invention are recognizable through sequence comparison to other known polynucleotides, and can be identified through use of alignment programs made available in public sequence databases.

The invention also provides fragment polynucleotides that are conserved
20 in one or more polynucleotides encoding members of the PDE10 family of polypeptides. Such fragments include sequences characteristic of the family of PDE10 polynucleotides, and are also referred to as "signature sequences." The conserved signature sequences are readily discernable following simple sequence comparison of polynucleotides encoding members of the PDE10 family. Fragments of the invention can be labeled in a manner that
25 permits their detection, and radioactive and non-radioactive labeling are comprehended.

Fragment polynucleotides are particularly useful as probes for detection of full length or other fragment PDE10 polynucleotides. One or more fragment polynucleotides can be included in kits that are used to detect the presence of a polynucleotide encoding PDE10, or used to detect variations in a polynucleotide sequence
30 encoding PDE10.

-8-

The invention further embraces species homologs, preferably mammalian, of the human PDE10 DNA. The polynucleotide sequence information provided by the invention makes possible the identification and isolation of polynucleotides encoding related mammalian PDE10 molecules by well known techniques including Southern and/or Northern hybridization, and polymerase chain reaction (PCR). Examples of related polynucleotides include human and non-human genomic sequences, including allelic variants, as well as polynucleotides encoding polypeptides homologous to PDE10 and structurally related polypeptides sharing one or more biological, immunological, and/or physical properties of PDE10.

The invention also embraces DNA sequences encoding PDE10 species which hybridize under moderately stringent conditions to the non-coding strands, or complements, of the polynucleotide in any one of SEQ ID NOs: 1, 18, 20, and 22. DNA sequences encoding PDE10 polypeptides which would hybridize thereto but for the redundancy of the genetic code are contemplated by the invention. Exemplary moderate hybridization conditions are as follows: hybridization at 65°C in 3X SSC, 0.1% Sarkosyl, and 20 mM sodium phosphate, pH 6.8, and washing at 65°C in 2X SSC with 0.1% SDS. Exemplary high stringency conditions would include a final wash in 0.2X SSC/0.1% SDS, at 65°C to 75°C. It is understood in the art that conditions of equivalent stringency can be achieved through variation of temperature and buffer, or salt concentration as described Ausebel, *et al.* (Eds.), Protocols in Molecular Biology, John Wiley & Sons (1994), pp. 6.0.3 to 6.4.10. Modifications in hybridization conditions can be empirically determined or precisely calculated based on the length and the percentage of guanosine/cytosine (GC) base pairing of the probe. The hybridization conditions can be calculated as described in Sambrook, *et al.*, (Eds.), Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory Press: Cold Spring Harbor, New York (1989), pp. 9.47 to 9.51.

Autonomously replicating recombinant expression constructions such as plasmid and viral DNA vectors incorporating PDE10 sequences are also provided. Expression constructs wherein PDE10-encoding polynucleotides are operatively-linked to an endogenous or exogenous expression control DNA sequence and a transcription terminator are also provided.

-9-

According to another aspect of the invention, host cells are provided, including procaryotic and eucaryotic cells, either stably or transiently transformed with DNA sequences of the invention in a manner which permits expression of PDE10 polypeptides of the invention. Expression systems of the invention include bacterial, yeast, fungal, viral, invertebrate, and mammalian cells systems. Host cells of the invention are a valuable source of immunogen for development of antibodies specifically immunoreactive with PDE10. Host cells of the invention are also conspicuously useful in methods for large scale production of PDE10 polypeptides wherein the cells are grown in a suitable culture medium and the desired polypeptide products are isolated from the cells or from the medium in which the cells are grown by, for example, immunoaffinity purification.

Knowledge of PDE10 DNA sequences allows for modification of cells to permit, or increase, expression of endogenous PDE10. Cells can be modified (*e.g.*, by homologous recombination) to provide increased PDE10 expression by replacing, in whole or in part, the naturally occurring PDE10 promoter with all or part of a heterologous promoter so that the cells express PDE10 at higher levels. The heterologous promoter is inserted in such a manner that it is operatively-linked to PDE10 encoding sequences. See, for example, PCT International Publication No. WO 94/12650, PCT International Publication No. WO 92/20808, and PCT International Publication No. WO 91/09955. The invention also comprehends that, in addition to heterologous promoter DNA, amplifiable marker DNA (*e.g.*, *ada*, *dhfr*, and the multifunctional CAD gene which encodes carbamyl phosphate synthase, aspartate transcarbamylase, and dihydroorotase) and/or intron DNA may be inserted along with the heterologous promoter DNA. If linked to the PDE10 coding sequence, amplification of the marker DNA by standard selection methods results in co-amplification of the PDE10 coding sequences in the cells.

The DNA sequence information provided by the present invention also makes possible the development through, *e.g.* homologous recombination or "knock-out" strategies [Capecchi, *Science* 244:1288-1292 (1989)], of animals that fail to express functional PDE10 or that express a variant of PDE10. Such animals are useful as models for studying the *in vivo* activities of PDE10 and modulators of PDE10.

The invention also provides purified and isolated mammalian PDE10 polypeptides as set out in SEQ ID NOs: 2, 19, 21, and 23. Presently preferred is a PDE10 polypeptide comprising the amino acid sequence set out in SEQ ID NO: 2. The invention embraces PDE10 polypeptides encoded by a DNA selected from the group consisting of:

5 a) the DNA sequence set out in SEQ ID NOs: 1, 18, 20, or 22; b) a DNA molecule which hybridizes under stringent conditions to the noncoding strand of the protein coding portion of (a); and c) a DNA molecule that would hybridize to the DNA of (a) but for the degeneracy of the genetic code. The invention also embraces polypeptide fragments of the sequences set out in SEQ ID NOs: 2, 19, 21, or 23 wherein the fragments maintain

10 biological or immunological properties of a PDE10 polypeptide. Preferred polypeptide fragments display antigenic properties unique to or specific for the PDE10 family of polypeptides. Fragments of the invention can be prepared by any the methods well known and routinely practiced in the art, having the desired biological and immunological properties.

15 The invention embraces polypeptides have at least 99%, at least 95%, at least 90%, at least 85%, at least 80%, at least 75%, at least 70%, at least 65%, at least 60%, at least 55% and at least 50% identity and/or homology to the preferred PDE10 polypeptide on the invention. Percent amino acid sequence "identity" with respect to the preferred polypeptide of the invention is defined herein as the percentage of amino acid

20 residues in the candidate sequence that are identical with the residues in the PDE10 sequence after aligning both sequences and introducing gaps, if necessary, to achieve the maximum percent sequence identity, and not considering any conservative substitutions as part of the sequence identity. Percent sequence "homology" with respect to the preferred polypeptide of the invention is defined herein as the percentage of amino acid

25 residues in the candidate sequence that are identical with the residues in the PDE10 sequence after aligning the sequences and introducing gaps, if necessary, to achieve the maximum percent sequence identity, and also considering any conservative substitutions as part of the sequence identity. Conservative substitutions can be defined as set out below.

30 PDE10 polypeptides of the invention may be isolated from natural cell sources or may be chemically synthesized, but are preferably produced by recombinant

-11-

procedures involving host cells of the invention. Use of various host cells is expected to provide for such post-translational modifications (*e.g.*, glycosylation, truncation, lipidation, and phosphorylation) as may be needed to confer optimal biological activity on recombinant expression products of the invention. PDE10 products of the invention may be full length polypeptides, biologically or immunologically active fragments, or variants thereof which retain specific PDE10 biological or immunological activity. Variants may comprise PDE10 polypeptide analogs wherein one or more of the specified (*i.e.*, naturally encoded) amino acids is deleted or replaced or wherein one or more non-specified amino acids are added: (1) without loss of one or more of the biological activities or immunological characteristics specific for PDE10; or (2) with specific disablement of a particular biological activity of PDE10.

Variant products of the invention include mature PDE10 products, *i.e.*, PDE10 products wherein leader or signal sequences are removed, and having additional, non-naturally occurring, amino terminal residues. PDE10 products having an additional methionine residue at position -1 (Met⁻¹-PDE10) are contemplated, as are PDE10 products having additional methionine and lysine residues at positions -2 and -1 (Met⁻²-Lys⁻¹-PDE10). Variants of these types are particularly useful for recombinant protein production in bacterial cell types.

The invention also embraces PDE10 variants having additional amino acid residues that result from use of specific expression systems. For example, use of commercially available vectors that express a desired polypeptide such as a glutathione-S-transferase (GST) fusion product provide the desired polypeptide having an additional glycine residue at position -1 as a result of cleavage of the GST component from the desired polypeptide. Variants which result from expression in other vector systems are also contemplated.

Variant polypeptides include those wherein conservative substitutions have been introduced by modification of polynucleotides encoding polypeptides of the invention. Conservative substitutions are recognized in the art to classify amino acids according to their related physical properties and can be defined as set out in Table I (from WO 97/09433, page 10, published March 13, 1997 (PCT/GB96/02197, filed 9/6/96).

-12-

Table I
Conservative Substitutions I

	<u>SIDE CHAIN CHARACTERISTIC</u>	<u>AMINO ACID</u>
5	Aliphatic	Non-polar
		G A P
		I L V
		Polar - uncharged
		C S T M
		N Q
		Polar - charged
		D E
		K R
	Aromatic	H F W Y
	Other	N Q D E

Alternatively, conservative amino acids can be grouped as defined in Lehninger,
10 [Biochemistry, Second Edition; Worth Publishers, Inc. NY:NY (1975), pp.71-77] as set
out in Table II.

Table II
Conservative Substitutions II

15	<u>SIDE CHAIN</u>	
	<u>CHARACTERISTIC</u>	<u>AMINO ACID</u>
	Non-polar (hydrophobic)	
	A. Aliphatic:	A L I V P
20	B. Aromatic:	F W
	C. Sulfur-containing:	M
	D. Borderline:	G
	Uncharged-polar	
	A. Hydroxyl:	S T Y
25	B. Amides:	N Q
	C. Sulfhydryl:	C
	D. Borderline:	G
	Positively Charged (Basic):	K R H
30	Negatively Charged (Acidic):	D E

The invention further embraces PDE10 products modified to include one
or more water soluble polymer attachments. Particularly preferred are PDE10 products

-13-

covalently modified with polyethylene glycol (PEG) subunits. Water soluble polymers may be bonded at specific positions, for example at the amino terminus of the PDE10 products, or randomly attached to one or more side chains of the polypeptide.

Also comprehended by the present invention are antibodies (*e.g.*,
5 monoclonal and polyclonal antibodies, single chain antibodies, chimeric antibodies, human antibodies CDR-grafted antibodies, or otherwise "humanized" antibodies, antigen binding antibody domains including Fab, Fab', F(ab')₂, F_v, or single variable domains, and the like) and other binding proteins specific for PDE10 products or fragments thereof. Specific binding proteins can be developed using isolated or recombinant PDE10
10 products, PDE10 variants, or cells expressing such products. The term "specific for" indicates that the variable regions of the antibodies recognize and bind PDE10 polypeptides exclusively (*i.e.*, able to distinguish PDE10 polypeptides from the superfamily of PDE polypeptides despite sequence identity, homology, or similarity found in the family of polypeptides), but may also interact with other proteins (for example, *S.*
15 *aureus* protein A or other antibodies in ELISA techniques) through interactions with sequences outside the variable region of the antibodies, and in particular, in the constant region of the molecule. Screening assays to determine binding specificity of an antibody of the invention are well known and routinely practiced in the art. For a comprehensive discussion of such assays, see Harlow *et al.* (eds), Antibodies: A Laboratory Manual, Cold
20 Spring Harbor Laboratory, Cold Spring Harbor, NY (1988), Chapter 6. Antibodies that recognize and bind fragments of the PDE10 polypeptides of the invention are also contemplated, provided that the antibodies are first and foremost specific for, as defined above, PDE10 polypeptides. As with antibodies that are specific for full length PDE10 polypeptides, antibodies of the invention that recognize PDE10 fragments are those which
25 can distinguish PDE10 polypeptides from the superfamily of PDE polypeptides despite inherent sequence identity, homology, or similarity found in the family of proteins.

Binding proteins are useful for purifying PDE10 products and detection or quantification of PDE10 products in fluid and tissue samples using known immunological procedures. Binding proteins are also manifestly useful in modulating (*i.e.*,
30 blocking, inhibiting or stimulating) biological activities of PDE10, especially those

-14-

activities involved in signal transduction. Anti-idiotypic antibodies specific for anti-PDE10 antibodies are also contemplated.

The scientific value of the information contributed through the disclosures of DNA and amino acid sequences of the present invention is manifest. As one series of examples, knowledge of the sequence of a cDNA for PDE10 makes possible through use of Southern hybridization or polymerase chain reaction (PCR) the identification of genomic DNA sequences encoding PDE10 and PDE10 expression control regulatory sequences such as promoters, operators, enhancers, repressors, and the like. DNA/DNA hybridization procedures carried out with DNA sequences of the invention under moderately to highly stringent conditions are likewise expected to allow the isolation of DNAs encoding allelic variants of PDE10; allelic variants are known in the art to include structurally related proteins sharing one or more of the biochemical and/or immunological properties specific to PDE10. Similarly, non-human species genes encoding proteins homologous to PDE10 can also be identified by Southern and/or PCR analysis and useful in animal models for PDE10-related disorders. As an alternative, complementation studies can be useful for identifying other human PDE10 products as well as non-human proteins, and DNAs encoding the proteins, sharing one or more biological properties of PDE10.

Polynucleotides of the invention are also useful in hybridization assays to detect the capacity of cells to express PDE10. Polynucleotides of the invention may also be the basis for diagnostic methods useful for identifying a genetic alteration(s) in a PDE10 locus that underlies a disease state or states.

The DNA and amino acid sequence information provided by the present invention also makes possible the systematic analysis of the structure and function of PDE10s. DNA and amino acid sequence information for PDE10 also permits identification of binding partner compounds with which a PDE10 polypeptide or polynucleotide will interact. Binding partner compounds include proteins and non-protein compounds such as small molecules. Agents that modulate (*i.e.*, increase, decrease, or block) PDE10 activity or expression may be identified by incubating a putative modulator with a PDE10 polypeptide or polynucleotide and determining the effect of the putative modulator on PDE10 phosphodiesterase activity or expression. The selectivity of a compound that modulates the activity of the PDE10 can be evaluated by comparing its

-15-

binding activity on the PDE10 to its activity on other PDE enzymes. Cell based methods, such as di-hybrid assays to identify DNAs encoding binding compounds and split hybrid assays to identify inhibitors of PDE10 polypeptide interaction with a known binding polypeptide, as well as *in vitro* methods, including assays wherein a PDE10 polypeptide, PDE10 polynucleotide, or a binding partner are immobilized, and solution assays are contemplated under the invention.

Selective modulators may include, for example, antibodies and other proteins or peptides which specifically bind to a PDE10 polypeptide or a PDE10-encoding nucleic acid, oligonucleotides which specifically bind to a PDE10 polypeptide or a PDE10 gene sequence, and other non-peptide compounds (e.g., isolated or synthetic organic and inorganic molecules) which specifically react with a PDE10 polypeptide or underlying nucleic acid. Mutant PDE10 polypeptides which affect the enzymatic activity or cellular localization of the wild-type PDE10 polypeptides are also contemplated by the invention. Presently preferred targets for the development of selective modulators include, for example: (1) regions of the PDE10 polypeptide which contact other proteins and/or localize the PDE10 polypeptide within a cell, (2) regions of the PDE10 polypeptide which bind substrate, (3) cyclic nucleotide-binding site(s) of the PDE10 polypeptide, (4) phosphorylation site(s) of the PDE10 polypeptide and (5) regions of the PDE10 polypeptide which are involved in multimerization of PDE10 subunits. Still other selective modulators include those that recognize specific PDE10 encoding and regulatory polynucleotide sequences. Modulators of PDE10 activity may be therapeutically useful in treatment of a wide range of diseases and physiological conditions in which PDE activity is known to be involved.

PDE10 polypeptides of the invention are particularly amenable to use in high throughput screening assays to identify binding partners, and preferably modulators. Cell based assays are contemplated, including yeast based assay systems as well as mammalian cell expression systems as described in Jayawickreme and Kost, *Curr. Opin. Biotechnol.* 8:629-634 (1997). Alternatively, automated and minaturized high throughput screening (HTS) assays, such as high density free format high density screening, as described in Houston and Banks, *Curr. Opin. Biotechnol.* 8:734-740 (1997). Combinatorial libraries are particularly useful in high throughput screening assays.

-16-

There are a number of different libraries used for the identification of small molecule modulators, including, (1) chemical libraries, (2) natural product libraries, and (3) combinatorial libraries comprised of random peptides, oligonucleotides or organic molecules.

5 Chemical libraries consist of structural analogs of known compounds or compounds that are identified as "hits" or "leads" via natural product screening. Natural product libraries are collections of microorganisms, animals, plants, or marine organisms which are used to create mixtures for screening by: (1) fermentation and extraction of
10 broths from soil, plant or marine microorganisms or (2) extraction of plants or marine organisms. Natural product libraries include polyketides, non-ribosomal peptides, and variants (non-naturally occurring) variants thereof. For a review, see *Science* 282:63-68 (1998). Combinatorial libraries are composed of large numbers of peptides, oligonucleotides, or organic compounds as a mixture. They are relatively easy to prepare by traditional automated synthesis methods, PCR, cloning or proprietary synthetic
15 methods. Of particular interest are peptide and oligonucleotide combinatorial libraries. Still other libraries of interest include protein, peptidomimetic, multiparallel synthetic collection, recombinatorial, and polypeptide libraries. For a review of combinatorial chemistry and libraries created therefrom, see Myers, *Curr. Opin. Biotechnol.* 8:701-707 (1997). Identification of modulators through use of the various libraries described herein
20 permits modification of the candidate "hit" (or "lead") to optimize the capacity of the "hit" to modulate activity.

 Also made available by the invention are anti-sense polynucleotides which recognize and hybridize to polynucleotides encoding PDE10. Full length and fragment anti-sense polynucleotides are provided. The worker of ordinary skill will appreciate that
25 fragment anti-sense molecules of the invention include (i) those which specifically recognize and hybridize to PDE10 RNA (as determined by sequence comparison of DNA encoding PDE10 to DNA encoding other known molecules) as well as (ii) those which recognize and hybridize to RNA encoding variants in the PDE10 family of proteins. Antisense polynucleotides that hybridize to RNA encoding other members of the PDE10
30 family of proteins are also identifiable through sequence comparison to identify characteristic, or signature, sequences for the family of molecules. Anti-sense

-17-

polynucleotides are particularly relevant to regulating expression of PDE10 by those cells expressing PDE10 mRNA.

Antisense nucleic acids (preferably 10 to 20 base pair oligonucleotides) capable of specifically binding to PDE10 expression control sequences or PDE10 RNA are introduced into cells (e.g., by a viral vector or colloidal dispersion system such as a liposome). The antisense nucleic acid binds to the PDE10 target nucleotide sequence in the cell and prevents transcription or translation of the target sequence. Phosphorothioate and methylphosphonate antisense oligonucleotides are specifically contemplated for therapeutic use according to the invention. The antisense oligonucleotides may be further modified by poly-L-lysine, transferrin polylysine, or cholesterol moieties at the 5' end.

The invention further comprehends methods to modulate PDE10 expression through use of ribozymes. For a review, see Gibson and Shillito, *Mol. Biotech.* 7:125-137 (1997). Ribozyme technology can be utilized to inhibit translation of PDE10 mRNA in a sequence specific manner through (i) the hybridization of a complementary RNA to a target mRNA and (ii) cleavage of the hybridized mRNA through nuclease activity inherent to the complementary strand. Ribozymes can be identified by empirical methods but more preferably are specifically designed based on accessible sites on the target mRNA [Bramlage, *et al.*, *Trends in Biotech* 16:434-438 (1998).] Delivery of ribozymes to target cells can be accomplished using either exogenous or endogenous delivery techniques well known and routinely practiced in the art. Exogenous delivery methods can include use of targeting liposomes or direct local injection. Endogenous methods include use of viral vectors and non-viral plasmids.

Ribozymes can specifically modulate expression of PDE10 when designed to be complementary to regions unique to a polynucleotide encoding PDE10. "Specifically modulate" therefore is intended to mean that ribozymes of the invention recognize only a polynucleotide encoding PDE10. Similarly, ribozymes can be designed to modulate expression of all or some of the PDE10 family of proteins. Ribozymes of this type are designed to recognize polynucleotide sequences conserved in all or some of the polynucleotides which encode the family of proteins.

The invention further embraces methods to modulate transcription of PDE10 through use of oligonucleotide-directed triplet helix formation. For a review, see

-18-

Lavrovsky, *et al.*, *Biochem. Mol. Med.* 62:11-22 (1997). Triplet helix formation is accomplished using sequence specific oligonucleotides which hybridize to double stranded DNA in the major groove as defined in the Watson-Crick model. Hybridization of a sequence specific oligonucleotide can thereafter modulate activity of DNA-binding proteins, including, for example, transcription factors and polymerases. Preferred target sequences for hybridization include promoter and enhancer regions to permit transcriptional regulation of PDE10 expression. Oligonucleotides which are capable of triplet helix formation are also useful for site-specific covalent modification of target DNA sequences. Oligonucleotides useful for covalent modification are coupled to various DNA damaging agents as described in Lavrovsky, *et al.* [*supra*].

The invention comprehends mutations in the PDE10 gene that result in loss of normal function of the PDE10 gene product and underlie human disease states in which failure of the PDE10 is involved. Gene therapy to restore PDE10 activity would thus be indicated in treating those disease states. Delivery of a functional PDE10 gene to appropriate cells is effected *ex vivo*, *in situ*, or *in vivo* by use of vectors, and more particularly viral vectors (*e.g.*, adenovirus, adeno-associated virus, or a retrovirus), or *ex vivo* by use of physical DNA transfer methods (*e.g.*, liposomes or chemical treatments). See, for example, Anderson, *Nature*, supplement to vol. 392, no. 6679, pp.25-20 (1998). For additional reviews of gene therapy technology see Friedmann, *Science*, 244: 1275-1281 (1989); Verma, *Scientific American*: 68-84 (1990); and Miller, *Nature*, 357: 455-460 (1992). Alternatively, it is contemplated that in other human disease states, preventing the expression of or inhibiting the activity of PDE10 will be useful in treating the disease states. It is contemplated that antisense therapy or gene therapy could be applied to negatively regulate the expression of PDE10.

Identification of modulators of PDE10 expression and/or biological activity provides methods to treat disease states that arise from aberrant PDE10 activity. Modulators may be prepared in compositions for administration, and preferably include one or more pharmaceutically acceptable carriers, such as pharmaceutically acceptable (*i.e.*, sterile and non-toxic) liquid, semisolid, or solid diluents that serve as pharmaceutical vehicles, excipients, or media. Any diluent known in the art may be used. Exemplary diluents include, but are not limited to, polyoxyethylene sorbitan monolaurate, magnesium

-19-

stearate, methyl- and propylhydroxybenzoate, talc, alginates, starches, lactose, sucrose, dextrose, sorbitol, mannitol, gum acacia, calcium phosphate, mineral oil, cocoa butter, and oil of theobroma. The modulator compositions can be packaged in forms convenient for delivery. The compositions can be enclosed within a capsule, sachet, cachet, gelatin, paper, or other container. These delivery forms are preferred when compatible with entry of the composition into the recipient organism and, particularly, when the composition is being delivered in unit dose form. The dosage units can be packaged, *e.g.*, in tablets, capsules, suppositories or cachets. The compositions may be introduced into the subject by any conventional method including, *e.g.*, by intravenous, intradermal, intramuscular, intramammary, intraperitoneal, or subcutaneous injection; by oral, sublingual, nasal, anal, vaginal, or transdermal delivery; or by surgical implantation, *e.g.*, embedded under the splenic capsule or in the cornea. The treatment may consist of a single dose or a plurality of doses over a period of time.

The invention also embraces use of a PDE10 polypeptide, a PDE10 polynucleotide, or a binding partner thereof in production of a medicament for treatment of a PDE10-related biological disorder.

The present invention is illustrated by the following examples relating to the isolation of a polynucleotide encoding a PDE10 polypeptide and expression thereof. Example 1 describes identification of an EST encoding a partial PDE10 polypeptide and isolation of a full length PDE10-encoding clone. Example 2 relates to Northern blot analysis of PDE10 expression. Example 3 addresses chromosome mapping of PDE10. Example 4 describes expression and characterization of a recombinant PDE10 polypeptide. Example 5 describes production of anti-PDE10 antibodies. Example 6 provides an analysis of PDE10 expression using *in situ* hybridization. Example 7 relates to high throughput screening to identify inhibitors of PDE10.

Example 1

Identification of an EST Related to a Human PDE and Isolation of a Full Length PDE10-encoding Polynucleotide

Using the sequences of known human, 3', 5' cyclic nucleotide phosphodiesterases, a search of the National Center for Biotechnology Information (NCBI) Expressed Sequence Tags (EST) database was undertaken in order to identify

-20-

cDNA fragments that could potentially be useful for the identification of novel phosphodiesterase (PDE) genes. This database contains DNA sequences representing one or both ends of cDNAs collected from a variety of tissue sources. A single sequencing run is performed on one or both ends of the cDNA and the quality of the DNA sequence varies tremendously. At the time the PDE searches were performed, the EST sequence database contained more than 600,000 cDNA sequences from a variety of organisms.

The search for novel PDE sequences included three steps. First, the BLASTN program available through NCBI was used to identify DNA sequences in the EST sequence database with homology to cDNA sequences encoding known human PDEs. The program compares a nucleotide query sequence against a nucleotide sequence database. The cDNA sequences of the fifteen known human PDEs were submitted and fifteen BLASTN searches were performed; the query PDE sequences included PDE1A3 [Loughney, *et al.*, *J. Biol. Chem.* 271:796-806 (1996)], PDE1B1 [Yu, *et al.*, *Cell Signaling*, 9:519-529 (1997)], PDE1C2 [Loughney, *et al.*, *J. Biol. Chem.* 271:796-806 (1996)], PDE2A3 [Rosman, *et al.*, *Gene* 191:89-95 (1997)], PDE3A [Meacci, *et al.*, *Proc. Natl. Acad. Sci. (USA)* 89:3721-3725 (1992)], PDE3B [Miki *et al.*, *Genomics* 36:476-485 (1996)], PDE4A5 [Bolger, *et al.*, *Mol. Cell. Biol.* 13:6558-6571 (1993)], PDE4B2 [Bolger, *et al.*, *Mol. Cell. Biol.* 13:6558-6571 (1993)], PDE4C [Bolger, *et al.*, *Mol. Cell. Biol.* 13:6558-6571 (1993)], PDE4D1 [Bolger, *et al.*, *Biochem. J.* 328:539-548 (1997)] and PDE4D3 [Bolger, *et al.*, *Mol. Cell. Biol.* 13:6558-6571 (1993)], PDE5A, PDE6A [Pittler, *et al.*, *Genomics* 6:272-283 (1990)], PDE6B [Collins, *et al.*, *Genomics* 13:698-704 (1992)], PDE6C [Piriev, *et al.*, *Genomics* 28:429-435 (1995)], and PDE7A1 [Michaeli, *et al.*, *J. Biol. Chem.* 17:12925-12932 (1993)]. The BLASTN results were examined and EST sequences that were judged as corresponding to each of the fifteen known PDE cDNAs were identified and collected into a table. The PDE6A and PDE6B sequences used as queries were truncated at 3' end (removing a portion of the 3' untranslated region) due to the presence of repetitive elements in the 3' untranslated region of the cDNAs.

Secondly, the NCBI TBLASTN program was used to examine the homology between the protein sequence of the fifteen known human PDEs (as above) and the six different possible proteins encoded by each of the EST DNA sequences. In this

-21-

search, the EST sequences are translated in the six possible reading frames and the amino acid sequences generated are compared to the query PDE amino acid sequences. Sequences identified as homologous at the amino acid level were examined and any EST sequences positively identified as corresponding to a known PDE during the BLASTN search described above were discarded.

The third step of the search involved analyzing the sequences that were not known PDEs. These amino acid sequences were homologous to a known PDE but were not identified as one of the 15 known PDE genes during the BLASTN searches.

The initial BLAST searches identified three EST sequences, designated X88347 (SEQ ID NO: 3), X88467 (SEQ ID NO: 4), and X88465 (SEQ ID NO: 5), that were obtained from an exon trapping experiment using chromosome 21 genomic DNA and found to encode an amino acid sequence having homology to the catalytic region of one or more of the PDE query sequences. X88347 showed homology with the amino acid sequences of PDE1A, 1B, 1C, 3A, 3B, 4A, 4B and 4D; X88467 showed homology to PDE1A, 1B, 1C, 4A, 4B, 4C, and D4; and X88465 was homologous to PDE1A and 1B amino acid sequences. At the 5' terminus, EST X88465 was 58 nucleotides shorter than was X88467 and was not considered further.

When X88347 was translated from nucleotides 1-222 and the resultant protein was compared to PDE1A, the two proteins were the same at 23 of 51 amino acid positions (45% identity). When X88467 was translated from nucleotide 3 to 155 and the resultant protein compared to PDE1A, 15 of 36 amino acids were the same (42% identity). Because ESTs X88347 and X88467 showed homology to two different regions of the catalytic region of PDE1A, it seemed possible that they represented two different exons from a novel PDE gene.

X88347 was used as a query in a BLASTN search of the NCBI EST database. In addition to itself, X88347 identified three other human EST sequences with high enough homology to suggest the sequences were derived from the same gene. EST R00718 (SEQ ID NO: 6) showed 91% identity to X88347. R00719 (SEQ ID NO: 7) represented the 3'-end of the same cDNA as R00718. R45187 (SEQ ID NO: 8) showed 88% identity to X88347. Two mouse cDNAs were also identified; W82786 (SEQ ID NO: 9)(91% identity) and W10517 (SEQ ID NO: 10) appeared to represent the mouse

-22-

homolog of X88347. A BLASTN search using W10517 as probe identified another sequence H90802 (SEQ ID NO: 11), which appeared to represent another human EST that may be part of the human PDE gene. The several human cDNAs were not identical to each other, and the quality of the sequencing was poor. The cDNA represented by the
5 R00719 and R00718 EST sequences was obtained from the American Type Culture Collection (Rockville, MD) which maintains and makes publicly available deposits of ESTs identified and sequenced by I.M.A.G.E., Lawrence Livermore National Laboratory, (Livermore, CA). The cDNA had been isolated from a fetal liver and spleen library and mapped to chromosome 21.

10 R00718/9 was sequenced upon receipt and found to be consistent with the EST database sequence. The polynucleotide and amino acids sequences for R00718/9 are set out in SEQ ID Nos: 12 and 13, respectively. The R00718/9 clone contained a 0.6 kb insert with a poly A tail at the 3'-end. The open reading frame encoded a protein with homology to other PDEs but did not extend to the 5' end of the cDNA. Beginning at
15 amino acid position 9, a QSDRE sequence was found. Corresponding D and E residues were found within all of the query sequences. Query sequences also included a conserved E(F/Y) sequence located amino terminal to the conserved D and E residues, but this sequence was not found in EST R00718/9. Instead, the EST contained eight amino acids followed by a stop codon. The R00718/9 cDNA appeared to diverge from the PDE query
20 sequences in the catalytic region and the open reading frame was not maintained. The disrupted open reading frame may suggest the presence of an intron that had not been removed or that the R00718/9 sequence was joined to some unidentified extraneous polynucleotide sequence. The gene represented by R00718/9 was designated PDE10.

In order to identify additional PDE10 sequences, a probe was generated
25 based on the PDE10 sequence and used to screen cDNA libraries. First, two primers, R71S100R (SEQ ID NO: 14) and R71A521H (SEQ ID NO: 15) were synthesized for use in PCR to amplify a 420 nucleotide portion of the R00718/9 DNA fragment (nucleotides 130 to 550). Primer R71S100R generated an *EcoRI* restriction site in the amplification product (underlined below) and primer R71A521H generated a *HindIII* site (also
30 underlined below). The PCR fragment was designed to include the region of R00718/9 homologous to other PDEs, but not the poly A tail.

-23-

R71S100R

(SEQ ID NO: 14)

AGTCGAATTCACCGTGAGAAGTCAGAAG

R71A521H

(SEQ ID NO: 15)

GTCAAAGCTTACATGGTCTTGTGGTGCC

5

The PCR reaction contained 50 pg R00719 cDNA, 10 ng/μl each primer, 0.2 mM dNTP, 1X PCR buffer (Perkin-Elmer), 2 mM MgCl₂, and 1.25 U *Taq* polymerase (Perkin-Elmer). The reaction was first maintained at 94°C for four minutes, after which thirty cycles of one minute 94°C, two minutes 50°C, and four minutes at 72°C were performed. The PCR fragment was purified using low melting point agarose gel electrophoresis.

For library screening, the PCR fragment was labeled with ³²P with a random priming kit (Boehringer Mannheim) according to manufacturer's instructions and used to screen 10⁶ cDNAs from a human heart cDNA library (Stratagene, La Jolla, CA), 5 x 10⁵ cDNAs from a human hippocampal cDNA library (Clontech, Palo Alto, CA), and 7.5 x 10⁵ cDNAs from a human fetal brain cDNA library (Stratagene). Hybridization was carried out overnight in buffer containing 3X SSC, 0.1% Sarkosyl, 20 mM sodium phosphate, pH 6.8, 10X Denhardt's solution, and 50 μg/ml salmon sperm DNA at 65°C. Eleven positives were obtained from the fetal brain library and three from the hippocampal library. Partial sequencing led to the selection of one, FB79c, for further characterization. The polynucleotide and deduced amino acid sequences for FB79c are set out in SEQ ID NOs: 16 and 17, respectively.

FB79c contained a 1.3 kb insert; the 3' end of FB79c extended further than that of R00718/9 and contained 12 adenosine residues of the poly A tail of R00718/9, an *Eco*RI site (GGAATTC), an additional fifty-nine nucleotides and a poly A sequence. At the 5' end, the sequence for FB79c differed from that of R00718/9 beginning at, and continuing 5' of, nucleotide 121 of R00718/9 (corresponding to nucleotide 744 of FB79c). The open reading frame in FB79c (encoding a protein with homology to the query PDEs) did not extend to the 5' end of the cDNA but ended in a stop codon at nucleotide 104.

A sequence within the FB79c DNA located upstream of the point of divergence from R00718/9 (but within the portion of the open reading frame with

-24-

homology to the other PDEs) was the region chosen for a probe in subsequent library screening. The isolated sequence selected was a 0.36 kb *EcoRV* fragment extending from nucleotide 308 to nucleotide 671 of FB79c and was used to screen 1.75×10^6 cDNAs from the fetal brain cDNA library (Stratagene). More than twenty cDNAs were identified and twelve were subjected to partial restriction mapping and DNA sequencing. More extensive sequencing on six of them led to the selection of clones FB76.2 and FB68.2 for complete sequencing. The polynucleotide and amino acid sequences for clone FB76.2 are set out in SEQ ID NOs: 18 and 19, respectively, and the polynucleotide and amino acid sequences for clone FB68.2 are set out in SEQ ID NOs: 20 and 21, respectively.

FB76.2 contained a 1.9 kb cDNA insert; the 3' end of the cDNA stopped one nucleotide short of the poly A tail found in clone FB79c and the sequence diverged from FB79c 5' of nucleotide 109 in clone FB79c (corresponding to nucleotide 715 in FB76.2). The open reading frame in the FB76.2 sequence that encoded a protein with homology to the PDE query sequences extended to the 5' end of the cDNA and the first methionine was encoded beginning at nucleotide 74. Assuming this residue to be the initiating methionine, the open reading frame of FB76.2 encoded a 533 amino acid protein with a predicted molecular weight of 61,708 Da.

Clone FB68.2 contained a 2 kb cDNA insert. At the 3' end, it extended to the poly A tail found in the FB79c sequence and the open reading frame extended to the 5' end of the cDNA. FB68.2 differed from FB76.2 by the presence of an additional internal 180 nucleotides (nucleotides 225 to 404 of FB68.2) following corresponding nucleotide 335 of FB76.2. Since the number of additional nucleotides in the FB68.2 insertion was divisible by three, it did not alter the reading frame as compared to FB76.2. The position of the insert with respect to maintaining the same reading frame suggested that the sequence might represent an exon found in some, but not all, PDE10 cDNAs. Alternatively, the additional sequence could be an intron that had not been removed from the FB68.2 cDNA.

Because the FB76.2 and FB68.2 differed from each other, additional PDE10 DNAs were obtained and analyzed to more accurately define the PDE10 nucleotide sequence. A 5' 0.3 kb *EcoRI* fragment of FB76.2 (corresponding to nucleotides 1 to 285) was isolated and used as a probe to screen 7.5×10^5 cDNAs from

-25-

the fetal brain cDNA library. Thirty seven positives were obtained, of which nineteen were first characterized with respect to fragment size (insert) that hybridized to the 0.3 kb *EcoRI* probe. Eight of the nineteen clones were subsequently characterized by partial sequencing. Two clones, FB93a and FB94a, contained 0.5 kb and 1.6 kb *EcoRI* fragments, respectively, that hybridized and were chosen for complete sequencing. The polynucleotide and amino acid sequences for clone FB93a are set out in SEQ ID NOs: 22 and 23, respectively, and the polynucleotide and amino acid sequences for clone FB94a are set out in SEQ ID NOs : 1 and 2, respectively.

FB93a contained a 1.5 kb insert which did not extend to the 3' end of FB76.2 but was ninety nucleotides longer than FB76.2 at the 5' end. The additional nucleotides encoded a stop codon beginning at position 47 which was in reading frame with the first methionine in FB76.2 described above (nucleotide 164 in FB93a). The position of the stop codon indicated the presence of a complete open reading frame and that FB76.2 probably represented a full length cDNA. Like FB76.2, FB93a did not contain the 180 nucleotide insert that was present in FB68.2.

FB94a contained a 1.5 kb cDNA insert and the 3' end extended almost 0.1 kb beyond the stop codon. The first methionine was encoded beginning at nucleotide 26, and assuming this residue to be the initiating methionine, FB94a encoded a 466 amino acid protein with a predicted molecular weight of 54,367 Da. FB94a differed from FB76.2 and FB93a by absence of a 149 nucleotide region which, if consistent with the sequences for FB76.2 and FB93a, would have been located after nucleotide 42. The absence of the 149 nucleotide sequence produced a putative initiator methionine that is in a different reading frame than that found in FB76.2 and FB93a. Like FB76.2 and FB93a, FB94a did not contain the 180 nucleotide region found in FB68.2.

A search of the EST data base with the FB94a and FB93a sequences identified yet another possible sequence for a PDE10 cDNA. The sequence of EST A158300 lacked both the 149 nucleotide and the 180 nucleotide sequences discussed above. In addition, A158300 also lacked 55 nucleotides immediately 3' to the 180 nucleotide region as found in the FB68.2 sequence. The open reading frame in A158300 extended to the 5' end and the first methionine corresponded to the same one used by FB76.2 and FB93a. The presence of the additional 55 nucleotide deletion from A158300

-26-

resulted in a different reading frame for the sequence between the site where the 149 nucleotides were deleted and the site where the 180 nucleotides were deleted.

The sequence information for PDE10 derived from these cDNA sequences can be summarized as follows. There is a 149 nucleotide sequence found in some clones (sequences FB76.2, FB93a, FB68.2) but not in all (sequences FB94a, A158300). The 149 nucleotide sequence is followed by a 44 nucleotide region that is present in all the PDE10 cDNAs analyzed to date. Following the 44 nucleotide region is a sequence of 235 nucleotides in length. The region can be present in its entirety (as found in the sequence for FB68.2) or without the first 180 nucleotides (as observed in sequences FB76.2 and FB93a). As still another alternative, the whole region can be removed (as found in the sequence for A158300). These possibilities predict six different mRNA structures, four of which have been isolated.

The presence or absence of the 149 nucleotide region may reflect the presence or absence of an exon, and the presence of all or some of the 235 nucleotide region may reflect alternative 3'splice acceptor site usage. As an alternative, it is also possible that the 235 nucleotide region represents two separate exons of 180 and 55 nucleotides in length. The presence or absence of the 149 nucleotide sequence alters the reading frame of the encoded protein as does the presence or absence of the 55 nucleotide sequence.

A number of single nucleotide differences have been observed in comparison of the various PDE10 cDNAs. R00718/9 has a cytosine at nucleotide position 155 whereas the other cDNAs have a thymidine at this position; this difference represented a silent change as proline is encoded by both sequences. R00718/9 also has a cytosine at position 161 whereas the other cDNAs have an adenosine at the same position; this difference also represented a silent change as both sequences encode alanine. FB94a has a guanosine at position 1383 whereas the other cDNAs have an adenosine at this position; as a consequence of the difference, FB94a encodes a glycine rather than a glutamic acid at that position. FB76.2 has an adenosine rather than a cytosine at position 1809; the difference does not effect an amino acid difference since the nucleotide position is located in the 3' untranslated region. FB79c also has one less adenosine in the string

-27-

of nucleotides between 1204 and 1215 than do the other cDNAs; this difference is also within the 3' untranslated region.

In comparison of a predicted PDE10 amino acid sequence with other known PDEs indicated that most, but not all, of the amino acids that are conserved among the query sequences were also found in PDE10. Comparison of the PDE10 catalytic region to PDE4A, PDE5A, and PDE7A revealed 32%, 30% and 34% identity, respectively.

Example 2 Northern Blot

In order to determine which cell and tissue types express PDE10, Northern blot analysis was carried out using a commercially prepared multi-tissue Northern blot (Clontech, Palo Alto, CA). The probe was a *EcoRI/BclI* fragment of the FB76.2 corresponding to nucleotides 1 to 883. Hybridization conditions were as previously described [Loughney et al., *supra*, (1996)].

Results indicated a 2.2 to 2.4 kb band which was strongest in kidney, present in heart, pancreas, and placenta, and weakest in brain, lung, skeletal muscle and liver. The band was fairly wide in placenta suggesting that it might contain a number of mRNAs of slightly different sizes.

Example 3 Chromosome Mapping

As mentioned above, the X88347, X88467, and X88465 ESTs were identified with an exon trapping procedure using DNA from chromosome 21 [Chen *et al.* 1996]. X88467 was identified as a new sequence with homology to a mouse calcium-, calmodulin-dependent phosphodiesterase Q01065 aa 52-103. XD88347 was identified to be the same as EST R00718 and similar to *Drosophila* cAMP dependent phosphodiesterase.P12252. Both of these sequences were placed in a category described as having strong homology to known protein sequences.

A search of the Sequence Tagged Sites (STS) database at NCBI revealed homology of the 3'-end of PDE10 to STS WI-13322 which has been mapped to region 220.72 cr. from the top of chromosome 21. The cDNA that this STS was derived from

-28-

begins at nucleotide 1899 of FB68.2, does not have the poly A tail and extends further 3' than FB68.2. It seems likely that this STS sequence represents a PDE10A transcript to which no poly (A⁺) tail has been added or a PDE10A transcript that uses an alternative site for poly (A⁺) addition. STS WI-13322 was placed on a Whitehead map of chromosome 21 near SGC35805, which is derived from the gene for the cystathionine beta-synthase (CBS). CBS has been mapped to chromosome 21 at 21q22.3 [Avramopoulos, et al, Hum. Genet. 90:566-568 (1993); Munke *et al.*, Hum. Genet. 42:550-559 (1988)].

A number of different genetic diseases map to this region of chromosome 21, for example, Down syndrome [Delabar, *et al.*, Eur. J. Hum. Genet. 1:114-124(1993)]. It is not clear that PDE10A falls within the Down syndrome critical region (DSCR) but it is possible that genes elsewhere on chromosome 21 also contribute to Down syndrome [Korenberg, et al., Proc. Natl. Acad. Sci. (USA) 91:4997-5001 (1994)]. As another example, a locus involved in bipolar affective disorder in some families has been mapped to 21q22.3 [Vallada, *et al.*, J.Affect. Disord. 41:217-221 (1996)]. Other examples include Knobloch syndrome, characterized by myopia and retinal degeneration and detachment [Sertie, *et al.*, Hum. Mol. Genet. 5:843-847 (1996)], and one or more genes responsible for congenital recessive deafness (DFNB8, DFNB10) [Veske, *et al.*, Hum. Mol. Genet. 5:165168 (1996); Bonne-Tamir, *et al.*, Am. J. Hum. Genet. 58:1254-1259 (1996)]. PDE10A may play a role in any or all of these disease states.

Example 4 Expression and Characterization of PDE10

The entire open reading frame of the PDE10 cDNA (clone FB94a) was placed into a yeast ADH vector including the alcohol dehydrogenase promoter. The construct was built in two steps.

The 5' end was generated using PCR and FB94a DNA as template. PCR was carried out using the 5' primer below (SEQ ID NO: 25) in combination with 3' primer R71A3 (SEQ ID NO: 26). The 5' primer includes an *Nco*I site (underlined in SEQ ID NO: 25 below) and the initiating methionine codon of FB94a is in bold. The 5' primer also adds a FLAG[®] epitope tag (Eastman Kodak, Rochester, NY) to the amino terminus

of the encoded protein; the FLAG[®] tag is an epitope (SEQ ID NO: 24) recognized by the monoclonal antibody M2 (Eastman Kodak).

5 FLAG® TAG (SEQ ID NO: 24)
Asp-Tyr-Lys-Asp-Asp-Asp-Lys

5' Primer (SEQ ID NO: 25)
TAGACCATGGACTACAAGGACGACGA-
TGACAAGATGGACGCATTCAGAAGCACT

R71A3 (SEQ ID NO: 26)
CGAGGAGTCAACTTCTTG

PCR was carried out using 5 μ l each primer (100 μ g/ml stock), 5 μ l 10X buffer (Perkin Elmer), 5 μ l 10X nucleotides (2 mM stock), 3 μ l $MgCl_2$ (25 mM stock), FB94a DNA, and 0.3 μ l *Taq* polymerase (Perkin Elmer) in a reaction volume of 50 μ l. After incubating the reaction mixture at 94°C for four minutes, 30 cycles of one minute at 94°C, two minutes at 50°C, and four minutes at 72°C were carried out. The PCR product was cleaved with *Nco*I and *Hinc*II and purified using agarose gel electrophoresis. The 3' sequence of PDE10 was isolated as a *Hinc*II/*Eco*RI fragment cleaved from FB94a and purified by agarose gel electrophoresis. The two fragments were combined and ligated into a *Nco*I/*Eco*RI-digested Bluescript[®] vector (Stratagene, La Jolla, CA), previously modified by the insertion of the ADH promoter previously removed from a YEpC-PADH2d vector [Price *et al. Meth. Enzymol.* 185:308-315 (1990)] as a *Sac*I/*Nco*I fragment, to generate plasmid PDE10-1. New junctions and sequence generated by PCR were verified by sequencing.

In the second step of plasmid construction, the *SacI/SaII* fragment from PDE10-1 containing the ADH promoter and PDE10 open reading frame was purified by two rounds of agarose gel electrophoresis and ligated into *SacI/SaII* cut YEPC-PADH2 vector.

Following transformation into BJ2-54, a yeast strain lacking endogenous PDE activity, a colony was selected, streaked out on SC-leu plates and a single colony carrying the PDE10 construct was chosen for further characterization. Following overnight growth in SC-leu media the culture was diluted 1:250 in fresh SC-leu and

-30-

grown overnight at 30°C until it reached a density of 10^7 cells/ml. The cells were collected by centrifugation, washed once with YEP 3% glycerol media, resuspended in YEP containing 3% glycerol, and grown at 30°C for another 24 hours. The cells were harvested by centrifugation, washed with water, and frozen at -70°C until use. Prior to use, an aliquot of the yeast extract was analyzed by SDS PAGE. A protein specific to yeast carrying the PDE10 expression construct that migrated on the SDS PAGE gels with the expected mobility (55.5 kDa) was observed by Coomassie blue staining.

Yeast cells (1×10^{10}) were thawed with 200 µg/ml each of pepstatin A, leupeptin, and aprotinin 1 mM DTT, and 20 µg/ml calpain inhibitors (I and II). Two hundred µl of glass beads (0.5 mm, acid washed) were added, and the mixture was vortexed for eight cycles of 30 seconds each. Samples were cooled for 4.5 minutes at 4°C between cycles. After lysis, 0.8 ml lysis buffer was added, the lysate separated from the beads, and the lysate centrifuged for 30 minutes at 100,000 x g in a Beckman TL-100 tabletop centrifuge. The supernatant was aliquoted, frozen in dry ice/ethanol, and stored at -70°C.

Kinetic assays were performed on a BIOMEK[®] 1000 programmable robotic station (Beckman Instruments). The range of final substrate concentration was 0.2 to 1000 µM for cAMP and 0.6-2000 nM for cGMP. The highest nucleotide concentration contained 1 to 1.5 million Cerenkov counts of ³²P-labeled substrate per assay. The enzyme preparation was initially diluted 1:500 (cAMP as substrate) or 1:50,000 (cGMP as substrate). The enzyme dilution buffer consisted of 25 mM Tris-HCl pH 8.9, 5 µM ZnSO₄, 5 mM MgCl₂, 0.1 mM DTT, 100 mM NaCl and 0.1 mg/ml BSA (Calbiochem; fatty acid free). Activity at each substrate concentration was derived from a linear fit of successive four-fold enzyme dilutions across the plate.

Assays were performed at 30°C for 15 minutes. After 12 minutes, 5 µl snake venom from *Crotalus atrox* (15 mg/ml protein) was added to each reaction. Assays were stopped by addition 200 µl of charcoal suspension (25 mg/ml activated charcoal in 0.1 M monobasic potassium phosphate). The plate was centrifuged at 2600 rpm, and 200 µl of each supernatant was transferred into Microbeta[®] counting plates and counted on a WALLAC Microbeta[®] by Cerenkov counting. Data were evaluated with a predesigned

-31-

Microsoft Excel[®] Spreadsheet, and the kinetic parameters were fitted to a Michaelis-Menton model using the program Table Curve[®] from Jandel Scientific.

Results indicated that the K_m for cGMP hydrolysis was $5 (\pm 1)$ nM and the K_m for cAMP hydrolysis was $160 (\pm 30)$ μ M. In the extract, cGMP hydrolytic activity was determined to be $0.035 (\pm 0.01)$ μ mol/min/mg, while cAMP hydrolysis was measured to be $0.52 (\pm 0.06)$ μ mol/min/mg. Thus, although PDE10 had much greater affinity for cGMP, the V_{max} for cAMP was 15-fold greater.

In order to distinguish PDE10 from other PDE families, a panel of PDE inhibitors with activities against defined PDE families was tested for PDE10 inhibition using cAMP as a substrate. The results of the assay are set out in Table 1 below.

TABLE 1
PDE10 Inhibition with Isozyme-specific PDE Inhibitors

Inhibitor	Target Family	PDE10 IC_{50} (μ M)	Target Family IC_{50} (μ M)
SCH46642	PDE1	14	0.2^5
EHNA	PDE2	477	0.8^2
Cilostamide	PDE3	100	$0.04-0.9^3$
Rolipram	PDE4	529	$0.18-0.5^4$
DMPPO	PDE5	9	0.003^1
IBMX	non-specific	59	$2-20^1$

1. Coste and Grodin, *Biochem. Pharmacol.* 50:1577-1585 (1995).
2. Podzuweit, *et al. Cell. Signaling* 7:733-738 (1995)
3. Manganiello *et al.*, in Isoenzymes of Cyclic Nucleotide Phosphodiesterases, Beavo and Houslay (Eds.), John Wiley and Sons, Ltd. , pp. 87-116 (1990)
4. Bolger *et al.*, *Mol. Cell. Biol.* 13:6558-6571 (1993)
5. Ahn, *et al.*, Abstract from the 9th International Conference on Second Messengers and Phosphoproteins, Nashville, TN, 1995, p. 86.

-32-

The results further distinguish PDE10 from PDEs in families 1 through 5 in that specific inhibitors for enzymes in those families are significantly less effective in inhibiting PDE10.

5

Example 5 **Production of Anti-PDE10 Antibodies**

A GST fusion protein was produced in *E. coli* to provide an antigen for generation of monoclonal antibodies to PDE10. An *EcoRI* fragment from FB76.2 (nucleotides 280 through 1829 in SEQ ID NO: 18) was inserted into the *EcoRI* site of pGEX3X (Pharmacia) and the resultant construct was transformed in the *E. coli* strain XL1 Blue. A GST-PDE10 fusion protein including 464 amino acids from PDE10 was expressed from this construct following induction with IPTG. The fusion protein was isolated using SDS-PAGE, the band of appropriate size excised from the gel following staining with cold 0.4 M KCl, and the protein obtained from the acrylamide by electroelution. The elution product was dialyzed against PBS and concentrated using Centriprep 10 and Centricon columns (Amicon, Beverly MA) prior to being injected into mice.

On day 0, four Balb/c mice were pre-bled and immunized by subcutaneous injection with a panel of antigens including 30 µg/mouse GST-PDE10 fusion protein in complete Freund's adjuvant in 200 µl total volume. The same injections were repeated at weeks three and nine in incomplete Freund's adjuvant. Ten days after the last immunization, test bleeds were obtained and screened by antigen capture ELISA and Western analysis.

In the ELISA, Immulon[®] 4 plates (Dynex, Cambridge, Massachusetts) were coated at 4°C with 50 µl/well of a solution containing 2 µg/ml GST-PDE10 in 50 mM carbonate buffer, pH 9.6. Plates were blocked with 0.5% fish skin gelatin (Sigma) for 30 minutes and 50 µl serum diluted in PBS with 0.5% Tween[®] 20 (PBST) was added. Serum dilutions ranged from 1:100 to 1:102,400 and were obtained by a series of doubling dilutions. After incubation at 37°C for 30 minutes and washing three times with PBST, 50 µl of horseradish peroxidase-conjugated goat anti-mouse IgG(fc) antibody (Jackson) (diluted 1:10000 in PBST) was added. Plates were incubated as above and washed four times with PBST. Antibody was detected with addition of tetramethyl

-33-

benzidine (Sigma Chemical, St. Louis, Missouri) and the color reaction was stopped after five minutes with the addition of 50 μ l of 15% H_2SO_4 . Absorbance at 450 nm was measured on a plate reader.

For Western analysis, SDS-PAGE gels were run with approximately 10 μ g yeast PDE10 extract and approximately 200 ng of gel-purified GST-PDE10 and the proteins were transferred to Immobilon-PVDF. A standard enhanced chemiluminescence (ECL) Western blot protocol was performed using BioRad goat anti-mouse IgG horseradish peroxidase as the secondary antibody.

In preparation of hybridomas, splenocytes from mice giving a positive result from the ELISA and/or Western blotting protocols above, were fused to NS-1 cells in a ratio of 5:1 by standard methods using polyethylene glycol 1500 (Boehringer Mannheim) [Harlow and Lane, Antibodies, a Laboratory Manual, Cold Spring Harbor Laboratory, p.211 (1988)]. The fused cells were resuspended in 200 ml RPMI containing 15% FBS, 100 mM sodium hypoxanthine, 0.4 mM aminopterin, 16 mM thymidine (HAT) (Gibco), 25 units/ml IL-6 (Boehringer Mannheim) and 1.5×10^6 murine thymocytes/ml and dispensed into ten 96-well flat bottom tissue culture plates (Corning, United Kingdom) at 200 μ l/well. Cells were fed on days 2, 4, and 6 post fusion by aspirating approximately 100 μ l from each well with an 18 G needle (Becton Dickinson) and adding 100 μ l/well plating medium described above except containing 10 units/ml IL-6 and lacking thymocytes. On days 9 to 12, supernatants from the fusion wells were screened by antigen capture ELISA using GST and GST-PDE10 and by ECL Western analysis as described above.

A positive signal of the expected size was obtained on both lanes of the Western blot using mouse blood and monoclonal antibodies with reactivity to the yeast recombinant protein were obtained in the subsequent fusion.

Example 6

Analysis of PDE10 Expression by *in situ* Hybridization

Expression of PDE10 was examined in tissue sections by *in situ* hybridization as described below.

-34-

Preparation of probe

An *EcoRI/PstI* restriction enzyme fragment from the cDNA FB93a (corresponding to nucleotides 370 through 978 in SEQ ID NO: 22) was subcloned into a Bluescript[®] vector (Stratagene, La Jolla, CA) to generate an expression plasmid designated PDE10A3A. The plasmid was digested with *EcoRI* and transcribed with T3 polymerase to generate an antisense probe. A sense probe was generated by digestion the plasmid with *BamHI* and transcribing with T7 polymerase. The PDE10 templates were transcribed using a RNA Transcription kit (Stratagene, La Jolla, CA) in a reaction containing 5 µl of 5X transcription buffer (Stratagene), 30 mM DTT (Stratagene), 0.8 mM each ATP, CTP, GTP (10 mM (Stratagene), 40 U RNase Block II (Stratagene), 12.5 U T3 or T7 polymerase (Stratagene), and 300 ng linearized plasmid template, 50 µCi ³⁵S-UTP (greater than 1000 Ci/mmol, Amersham, Arlington Heights, IL). The mixture was incubated at 37°C for one hour after which the template DNA was removed by addition of 1 µl of RNase-free DNase I (Stratagene) and incubation for 15 minutes at 37°C. The probe was hydrolyzed to approximately 250 nucleotides in length to facilitate tissue penetration by adding 4 µl 1 M NaHCO₃ and 6 µl 1 M Na₂CO₃ for 22 minutes at 60°C and the reaction mixture was neutralized by addition of 25 µl of a solution containing 100 µl 3 M sodium acetate, 5 µl acetic acid (VWR, So. Plainfield, NJ), and 395 µl dH₂O. A Quick Spin G50 RNA column (5'-3' Inc., Boulder, CO) was prepared according to the manufacturer's suggested protocol. The probe was placed in the center of the column and the column centrifuged for four minutes at 1,000 rpm in a desk top centrifuge. The column flow-through was mixed with 50 µl dH₂O, 2 µl of a 10 mg/ml tRNA solution, 10 µl 3 M sodium acetate, and 200 µl 100% ethanol (VWR) and the resulting mixture was incubated at -20°C overnight. The probe solution was microfuged for 15 minutes at 4°C, the supernatant was removed, and the pellet was resuspended in 40 µl 1X TBE containing 1 µl of 0.1 M DTT. The probe was stored at -70°C until the *in situ* hybridization assay was performed.

Preparation of tissue samples and *in situ* hybridization

Tissues (National Disease Research Interchange, Philadelphia, PA and Cooperative Human Tissue Network, Philadelphia, PA) were sectioned at 6 µm and

-35-

placed on Superfrost Plus slides (VWR). Sections were fixed for 20 minutes at 4°C in 4% paraformaldehyde (Sigma, St. Louis, MO). The slides were rinsed in three changes of 1X calcium-, magnesium-free phosphate buffered saline (CMF-PBS), dehydrated with three successive washes with 70% ethanol, 95% ethanol and 100% ethanol, and dried for 30 minutes at room temperature. The slides were placed in 70% formamide (J.T. Baker) in 2X SSC for two minutes at 70°C, rinsed in 2X SSC at 4°C, dehydrated through 70%, 95% and 100% ethanol washes, and dried for 30 minutes at room temperature.

A prehybridization step was performed by placing the slides in an airtight box containing a piece of filter paper saturated with buffer containing 50% formamide (J.T. Baker) in 4X SSC. Each section was covered with 100 µl of rHB2 buffer consisting of 10% dextran sulfate (Sigma), 50% formamide (J.T. Baker, Phillipsburg, NJ), 100 mM DTT (Boehringer Mannheim, Indianapolis, IN), 0.3 M NaCl (Sigma), 20 mM Tris, pH 7.5, 5 mM EDTA (Sigma), and 1X Denhardt's solution (Sigma) and the slides were incubated at 42°C for two hours. The probe, as described above, was prepared by mixing 4 x 10⁵ cpm/tissue section with 5 µl of a 10 mg/ml tRNA solution per section and heating the mixture at 95°C for three minutes. Ice cold rHB2 buffer was added to bring the final volume to 20 µl/section. The probe-containing solution (20 µl/section) was added to 100 µl rHB2 buffer previously applied. The slides were incubated at 55°C for 12 to 16 hours. Following hybridization, the slides were washed once in 4X SSC containing 10 mM DTT for one hour at room temperature, once in 50% deionized formamide (J.T. Baker), 1X SSC, and 1 mM DTT for 40 minutes at 60°C, once in 2X SSC for 30 minutes at room temperature, and once in 0.1X SSC for 30 minutes at room temperature. The sections were dehydrated through 70%, 95%, and 100% ethanol washes and air dried for 30 minutes. The slides were dipped in Kodak NTB2 nuclear emulsion, dried for one to three hours at room temperature in the dark, and stored in the dark at 4°C with desiccant until time of development. The slides were developed in 4°C Kodak Dektol[®] developer for two minutes, dipped four times in 4°C dH₂O, and placed in 4°C Kodak fixer for ten minutes. The slides were rinsed in dH₂O and a standard hematoxylin and eosin (H&E) stain was performed as follows.

The slides were rinsed in dH₂O and stained with hematoxylin and eosin by transfer of the slides through a series of the following steps: five minutes in

-36-

formaldehyde/alcohol (100 ml formaldehyde, 900 ml 80% ethanol); three rinses in water for a total of two minutes; five minutes in 0.75% Harris hematoxylin (Sigma); three rinses in water for a total of two minutes; one dip in 1% HCl/50% ethanol; one rinse in water; four dips in 1% lithium carbonate; ten minutes in tap water; two minutes in 0.5% eosin (Sigma); three rinses in water for a total of two minutes; two minutes in 70% ethanol; three one-minute rinses in 95% ethanol; two one-minute rinses in 100% ethanol; and two two-minute rinses in xylene. Slides were mounted with cytooseal 60 (Stephens Scientific, Riverdale, NJ).

The signals obtained with an antisense PDE10 probe were compared to the control signals generated by a sense PDE10 probe and any signal specific to the antisense probe was assumed to represent PDE10 expression. PDE10 signal was detected throughout much of the cerebellum, with very strong signal in the Purkinje cells.

Example 7 **High Throughput Screening for PDE10 Inhibitors**

In an attempt to identify specific inhibitors, PDE10 was screened against a chemical library containing compounds of known structure. Initial screening was performed on pools of compounds (22 compounds per pool) with each compound present at 4.6 μ M. Pools which inhibited PDE10 activity by greater than 50% were selected and the individual compounds in the pool were screened at a concentration of 20 μ M. IC_{50} values were determined for compounds that inhibited enzyme activity.

An extract was prepared from *Saccharomyces cerevisiae* strain BJ2-54 (described in Example 4) lacking endogenous PDE activity and having PDE10 at an activity of 49 nmol cGMP hydrolyzed/min/ml with 32 nM cGMP. The extract was diluted 1:21,000-fold for use in the assay. Dilution buffer included 25 mM Tris, pH 8.0, 0.1 mM DTT, 5.0 mM $MgCl_2$, 100 mM NaCl, 5 μ M $ZnSO_4$ and 100 μ g/ml BSA. PDE assay buffer (5X) contained 200 mM Tris, pH 8.0, 5 mM EGTA, 25 mM $MgCl_2$ and 0.5 mg/ml BSA. Just prior to screening, 5X PDE assay buffer, deionized water, and 5'-nucleosidase (stock solution 15 mg/ml snake venom 5'-nucleosidase in 20 mM Tris, pH 8.0) were mixed at ratios of 4:4:1 to make Assay Reagent Mix.

A Packard MultiPROBE[®] was used to add 45 μ l of the Assay Reagent Mix and 20 μ l of the chemical compound pools. A BIOMEK[®] 1000 (See Example 4)

-37-

was used to add 20 μ l of PDE10 extract diluted as described above and 20 μ l ^{32}P -cGMP (ICN, specific activity 250 $\mu\text{Ci}/\text{mmol}$, diluted to 0.4 $\mu\text{Ci}/\text{ml}$, 16 nM, in deionized water). Final cGMP concentration in the assay was 0.08 $\mu\text{Ci}/\text{ml}$, 3.2 nM. Ten minutes after addition of ^{32}P -cGMP, 140 μ l of 25 mg/ml charcoal (in 0.1 M NaH_2PO_4) was added to stop the reaction. After a 20 minute incubation at room temperature, the assay plates were centrifuged for five minutes at 3,500 rpm in a Beckman GS-6R centrifuge. A BIOMEK[®] 1000 was used to transfer 140 μ l of the supernatant to a Wallac counting plate and Cerenkov radiation was measured in a Wallac MicroBeta Counter.

Several compounds that merit further investigation were found to inhibit enzyme activity.

Numerous modifications and variations in the invention as set forth in the above illustrative examples are expected to occur to those skilled in the art. Consequently only such limitations as appear in the appended claims should be placed on the invention.

-38-

What is claimed is:

1. A purified and isolated PDE10 polypeptide.
2. The polypeptide according to claim 1 comprising the amino acid sequence selected from the group consisting of SEQ ID NO: 2, SEQ ID NO: 18, SEQ ID NO: 20 and SEQ ID NO: 22.
3. A polynucleotide encoding the polypeptide according to claim 1 or 2.
4. The polynucleotide according to claim 3 comprising the sequence set forth in SEQ ID NO: 1.
5. A polynucleotide encoding a human PDE10 polypeptide selected from the group consisting of:
 - a) the polynucleotide according to claim 4;
 - b) a DNA which hybridizes under moderately stringent conditions to the non-coding strand of the polynucleotide of (a); and
 - c) a DNA which would hybridize to the non-coding strand of the polynucleotide of (a) but for the redundancy of the genetic code.
6. The polynucleotide of claim 5 comprising the polynucleotide sequence set out in SEQ ID NO: 18.
7. The polynucleotide of claim 5 comprising the polynucleotide sequence set out in SEQ ID NO: 20.

-39-

8. The polynucleotide of claim 5 comprising the polynucleotide sequence set out in SEQ ID NO: 22.

9. The polynucleotide of claim 5 which is a DNA molecule.

10. The DNA of claim 9 which is a cDNA molecule.

11. The DNA of claim 9 which is a wholly or partially chemically synthesized DNA molecule.

12. A polynucleotide comprising the sequence set out in SEQ ID NO: 1 or a fragment thereof.

13. A polynucleotide comprising the sequence set out in SEQ ID NO: 18 or a fragment thereof.

14. A polynucleotide comprising the sequence set out in SEQ ID NO: 20 or a fragment thereof.

15. A polynucleotide comprising the sequence set out in SEQ ID NO: 22 or a fragment thereof.

16. An anti-sense polynucleotide which specifically hybridizes with the complement of the polynucleotide of claim 5.

17. A expression construct comprising the polynucleotide according to claim 5.

18. A host cell transformed or transfected with the expression construct according to claim 17.

-40-

19. A method for producing a PDE10 polypeptide comprising the steps of:
- a) growing the host cell according to claim 18 under conditions appropriate for expression of the PDE10 polypeptide and
 - b) isolating the PDE10 polypeptide from the host cell or the medium of its growth.
20. An antibody specifically immunoreactive with the polypeptide according to claim 1 or 2.
21. The antibody according to claim 20 which is a monoclonal antibody.
22. A hybridoma which produces the antibody according to claim 21.
23. An anti-idiotypic antibody specifically immunoreactive with the antibody according to claim 21.
24. A method to identify a specific binding partner compound of the PDE10 polypeptide according to claim 1 or 2 comprising the steps of:
- a) contacting the PDE10 polypeptide with a compound under conditions which permit binding between the compound and the PDE10 polypeptide;
 - b) detecting binding of the compound to the PDE10 polypeptide; and
 - c) identifying the compound as a specific binding partner of the PDE10 polypeptide.

-41-

25. The method according to claim 24 wherein the specific binding partner modulates activity of the PDE10 polypeptide.

26. The method according to claim 25 wherein the compound inhibits activity of the PDE10 polypeptide.

27. The method according to claim 25 wherein the compound enhances activity of the PDE10 polypeptide.

28. A method to identify a specific binding partner compound of the PDE10 polynucleotide according to claim 5 comprising the steps of:

- a) contacting the PDE10 polynucleotide with a compound under conditions which permit binding between the compound and the PDE10 polynucleotide;
- b) detecting binding of the compound to the PDE10 polynucleotide; and
- c) identifying the compound as a specific binding partner of the PDE10 polynucleotide.

29. The method according to claim 28 wherein the specific binding partner modulates expression of a PDE10 polypeptide encoded by the PDE10 polynucleotide.

30. The method according to claim 29 wherein the compound inhibits expression of the PDE10 polypeptide.

31. The method according to claim 29 wherein the compound enhances expression of the PDE10 polypeptide.

32. A compound identified by the method according to claim 24 or 28.

-42-

33. A composition comprising the compound according to claim 32 and a pharmaceutically acceptable carrier.

- 1 -

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gac cgt gag aag tca gaa ggc ctt cct gtg gca ccg ttc atg gac cga Asp Arg Glu Lys Ser Glu Gly Leu Pro Val Ala Pro Phe Met Asp Arg 365 370 375	1156
gac aaa gtg acc aag gcc aca gcc cag att ggg ttc atc aag ttt gtc Asp Lys Val Thr Lys Ala Thr Ala Gln Ile Gly Phe Ile Lys Phe Val 380 385 390	1204

-4-

ctg atc cca atg ttt gaa aca gtg acc aag ctc ttc ccc atg gtt gag 1252
Leu Ile Pro Met Phe Glu Thr Val Thr Lys Leu Phe Pro Met Val Glu
395 400 405

gag atc atg ctg cag cca ctt tgg gaa tcc cga gat cgc tac gag gag 1300
Glu Ile Met Leu Gln Pro Leu Trp Glu Ser Arg Asp Arg Tyr Glu Glu
410 415 420 425

ctg aag cgg ata gat gac gcc atg aaa gag tta cag aag aag act gac 1348
Leu Lys Arg Ile Asp Asp Ala Met Lys Glu Leu Gln Lys Lys Thr Asp
430 435 440

agc ttg acg tct ggg gcc acc gag aag tcc aga ggg aga agc aga gat 1396
Ser Leu Thr Ser Gly Ala Thr Glu Lys Ser Arg Gly Arg Ser Arg Asp
445 450 455

gtg aaa aac agt gaa gga gac tgt gcc tgaggaaagc ggggggcgtg 1443
Val Lys Asn Ser Glu Gly Asp Cys Ala
460 465

gctgcagttc tggacgggct ggccgagctg cgcgggatcc ttgtgcaggg aagagctgcc 1503

ctgggcacct ggcaccacaa gaccatgttt tctaagaacc atttt 1548

<210> 2

<211> 466

<212> PRT

<213> Homo sapiens

<400> 2

Met Asp Ala Phe Arg Ser Thr Pro Tyr Lys Val Arg Pro Val Ala Ile
1 5 10 15

Lys Gln Leu Ser Glu Arg Glu Glu Leu Ile Gln Ser Val Leu Ala Gln
20 25 30

Val Ala Glu Gln Phe Ser Arg Ala Phe Lys Ile Asn Glu Leu Lys Ala
35 40 45

Glu Val Ala Asn His Leu Ala Val Leu Glu Lys Arg Val Glu Leu Glu
50 55 60

-5-

Gly Leu Lys Val Val Glu Ile Glu Lys Cys Lys Ser Asp Ile Lys Lys
 65 70 75 80

Met Arg Glu Glu Leu Ala Ala Arg Ser Ser Arg Thr Asn Cys Pro Cys
 85 90 95

Lys Tyr Ser Phe Leu Asp Asn His Lys Lys Leu Thr Pro Arg Arg Asp
 100 105 110

Val Pro Thr Tyr Pro Lys Tyr Leu Leu Ser Pro Glu Thr Ile Glu Ala
 115 120 125

Leu Arg Lys Pro Thr Phe Asp Val Trp Leu Trp Glu Pro Asn Glu Met
 130 135 140

Leu Ser Cys Leu Glu His Met Tyr His Asp Leu Gly Leu Val Arg Asp
 145 150 155 160

Phe Ser Ile Asn Pro Val Thr Leu Arg Arg Trp Leu Phe Cys Val His
 165 170 175

Asp Asn Tyr Arg Asn Asn Pro Phe His Asn Phe Arg His Cys Phe Cys
 180 185 190

Val Ala Gln Met Met Tyr Ser Met Val Trp Leu Cys Ser Leu Gln Glu
 195 200 205

Lys Phe Ser Gln Thr Asp Ile Leu Ile Leu Met Thr Ala Ala Ile Cys
 210 215 220

His Asp Leu Asp His Pro Gly Tyr Asn Asn Thr Tyr Gln Ile Asn Ala
 225 230 235 240

Arg Thr Glu Leu Ala Val Arg Tyr Asn Asp Ile Ser Pro Leu Glu Asn
 245 250 255

His His Cys Ala Val Ala Phe Gln Ile Leu Ala Glu Pro Glu Cys Asn
 260 265 270

Ile Phe Ser Asn Ile Pro Pro Asp Gly Phe Lys Gln Ile Arg Gln Gly
 275 280 285

-6-

Met Ile Thr Leu Ile Leu Ala Thr Asp Met Ala Arg His Ala Glu Ile
 290 295 300

Met Asp Ser Phe Lys Glu Lys Met Glu Asn Phe Asp Tyr Ser Asn Glu
 305 310 315 320

Glu His Met Thr Leu Leu Lys Met Ile Leu Ile Lys Cys Cys Asp Ile
 325 330 335

Ser Asn Glu Val Arg Pro Met Glu Val Ala Glu Pro Trp Val Asp Cys
 340 345 350

Leu Leu Glu Glu Tyr Phe Met Gln Ser Asp Arg Glu Lys Ser Glu Gly
 355 360 365

Leu Pro Val Ala Pro Phe Met Asp Arg Asp Lys Val Thr Lys Ala Thr
 370 375 380

Ala Gln Ile Gly Phe Ile Lys Phe Val Leu Ile Pro Met Phe Glu Thr
 385 390 395 400

Val Thr Lys Leu Phe Pro Met Val Glu Glu Ile Met Leu Gln Pro Leu
 405 410 415

Trp Glu Ser Arg Asp Arg Tyr Glu Glu Leu Lys Arg Ile Asp Asp Ala
 420 425 430

Met Lys Glu Leu Gln Lys Lys Thr Asp Ser Leu Thr Ser Gly Ala Thr
 435 440 445

Glu Lys Ser Arg Gly Arg Ser Arg Asp Val Lys Asn Ser Glu Gly Asp
 450 455 460

Cys Ala

465

<210> 3

<211> 225

<212> DNA

<213> Homo sapiens

- 7 -

<400> 3

agcgaccgtg agaagtcaga aggccttcct gtggaaccgt tcatggaccg agacaaagtg 60
accaaggcca cagcccagat tgggttcacg aagtttgccc tgatcccaat gtttgaaaca 120
gtgaccaagn tcttcccat ggttgaggag atcatgctgc agccactttg ggaatcccga 180
gatcgntacg aggagctgaa gcggnatagat gacgccatga aagag 225

<210> 4

<211> 158

<212> DNA

<213> Homo sapiens

<400> 4

gtaccagatc antgcccga cagagctggc ggtccgntac aatgacatct caccgttgga 60
gnaaccacca ctgcgccgtg gccttccaga tctcgcgga gcctgagtgn aacatcttct 120
ccaacatccc acctgatggg ttcaagcaga tccgacag 158

<210> 5

<211> 98

<212> DNA

<213> Homo sapiens

<400> 5

gagaacacca ctgngccgtg gncttccaga tctcgcgga gcctgagtgn aacatcttct 60
ccaacatccc acctgatggg ttcaagcaga tccgacag 98

<210> 6

<211> 418

<212> DNA

- 8 -

<213> Homo sapiens

<400> 6

nggttaactg gcgcattctg tctttctctg agaacagcga tctggttatg gggcatttct 60
gtctctaatag tcaactgtctg ctgcattccc tgcagagcga ccgtgagaag tcagaaggcc 120
ttcccgtggc cccgttcatg gaccgagaca aagtgaccaa ggccacagcc caggattggg 180
tttcatcaag tttgtctga tcccaatggt tgaaacagt accaagctct tccccatggg 240
ttgagggaga ttcattgttg cagccanttt ggggaatccc gaggattcgc tacgagggag 300
cttgaagcgg gattaggatg gacggccatg gaaaggagtt ttacaggaag gnaggatttg 360
acagttttga agttttgggg gggccaccga ggaagtccn ggaggaggag naggcaga 418

<210> 7

<211> 428

<212> DNA

<213> Homo sapiens

<400> 7

nagaaaaaag tgaacaaaat ggttcttaga aaacatggtc ttgtggtgcc aggtgcccag 60
ggagctcttc cctgcacaag gntcccgcgc antcgccag cccgtccaga actgcagcca 120
cgccccccgn ttctctcagg cacagtctcc ttactgttt ttacatctc tgcttctctc 180
tctggacttc tcggtggccc cagacgtcaa gctgtcagtc ttcttctgta actctttcat 240
gggcgtcatc tatccgcttc agtcctcgt aggcgatctc ggggattccc aaagtgggct 300
gcagcatgat ctctctcaac catggggggg aggagcttgg ggcactngtt ttcaaaaatt 360

-9-

gggggatcag gggacaaact ttgattggan cccatnttgg ggcttttggg cctttggggc 420

aatttttg 428

<210> 8

<211> 438

<212> DNA

<213> Homo sapiens

<400> 8

tttttttttt ttttttttgt atcagtgaac aaaatggttc ttagaaaaca tggctctgtg 60

gtncagggtg cccagggagc tcttccctgc acaaggancc cgcgcantcg gccagcccg 120

ccagaactgc agccacgccc cccgttttcc tcaggcacag tctccttcac tgtttttcac 180

atctctgntt ctctctctgg gantnttcgg tgggccccag aacgtcaagc tgtcagtntt 240

cttctgtaac tntttcatgg gcgtcatcta tccgtttcag cttcctcgta ggcgatnttg 300

gggattccca aagtgggctg gcagcatgga tcttcctcaa accatggggg gaaggagttt 360

gggtcaattn ttttcaaac attgggggnt cagggacaaa attttgatgg aaaccaatt 420

tgggggntgt gggccttg 438

<210> 9

<211> 262

<212> DNA

<213> Mus musculus

-10-

<400> 9

gagaattttg actacagcaa cgaggagcac ctgaccctgc tgaagatgat ttcataaaa 60
tgctgtgata tctccaatga agtccgtccc atggaggtgg cagaatcgtg ggtggactgt 120
ttactggaag aatattttat gcagagtgac cgtgagaagt ccgaagcctt cctgtggccc 180
cattcatgga ccgagacaaa gtgaccaaag caacagccca aattgggttc atcaagtttg 240
tcctgatccc aatgtttgaa ac 262

<210> 10

<211> 250

<212> DNA

<213> Mus musculus

<400> 10

gagaattttg actacagcaa cgaggagcac ctgaccctgc tgaagatgat ttcataaaa 60
tgctgtgata tctccaatga agtccgtccc atggaggtgg cagaatcgtg ggtggactgt 120
ttactggaag aatattttat gcagagtgac cgtgagaagt ccgaagcctt cctgtggccc 180
attcatggac cgagacaaag tgaccaaagc aacagccaaa ttgggttcat caagtttgtc 240
tgtccaatgt 250

<210> 11

<211> 459

<212> DNA

- 11 -

<213> Homo sapiens

<400> 11

attaatcttg gccactgaca tggcaagaca tgcagaaatt atggattctt tcaaagagaa 60

aatggagaat ttgactaca gcaacgagga gcacatgacc ctggtgagtg gcttattctg 120

cctgggtggg cagccaggcg gttgggctgg cgaanagggt catccatcca gctcacactg 180

gaagccaaga agctgaaatt attagtcttc ttggaacaag gtgtctataa atctggtttt 240

caaggtcag actcttacta ggaaagtccg ggcagggcct ccctcctgat gggctcctcct 300

tcatggtcag aggcagcatt ctcccattcc tccatctctt ttgggatttt gaaggagata 360

aagtgggggtg aaggccgtgc attctcgctc tgnntttcca gagaattaaa accagttttc 420

ccttgaaggc acagccccag cntggcattt tgaaagttg 459

<210> 12

<211> 599

<212> DNA

<213> Homo sapiens

<220>

<221> CDS

<222> (99)..(443)

<400> 12

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agaacagcga tctgggtatg gggcatttct gtctctaa tgt cac tgt ctg ctg cat 116
Cys His Cys Leu Leu His

1

5

- 12 -

tcc ctg cag agc gac cgt gag aag tca gaa ggc ctt ccc gtg gcc ccg 164
Ser Leu Gln Ser Asp Arg Glu Lys Ser Glu Gly Leu Pro Val Ala Pro
10 15 20

ttc atg gac cga gac aaa gtg acc aag gcc aca gcc cag att ggg ttc 212
 Phe Met Asp Arg Asp Lys Val Thr Lys Ala Thr Ala Gln Ile Gly Phe
 25 30 35

atc aag ttt gtc ctg atc cca atg ttt gaa aca gtg acc aag ctc ttc 260
Ile Lys Phe Val Leu Ile Pro Met Phe Glu Thr Val Thr Lys Leu Phe
40 45 50

ccc atg gtt gag gag atc atg ctg cag cca ctt tgg gaa tcc cga gat 308
Pro Met Val Glu Glu Ile Met Leu Gln Pro Leu Trp Glu Ser Arg Asp
55 60 65 70

cgc tac gag gag ctg aag cgg ata gat gac gcc atg aaa gag tta cag 356
 Arg Tyr Glu Glu Leu Lys Arg Ile Asp Asp Ala Met Lys Glu Leu Gln
 75 80 85

aag aag act gac agc ttg acg tct ggg gcc acc gag aag tcc aga gag 404
Lys Lys Thr Asp Ser Leu Thr Ser Gly Ala Thr Glu Lys Ser Arg Glu
90 95 100

aga agc aga gat gtg aaa aac agt gaa gga gac tgt gcc tgaggaaagc 453
Arg Ser Arg Asp Val Lys Asn Ser Glu Gly Asp Cys Ala
105 110 115

ggggggcgtg gctgcagttc tggacgggct ggccgagctg cgcgggatcc ttgtgcaggg 513

aagagctgcc ctgggcacct ggcaccacaa gaccatgttt tctaagaacc attttqttca 573

ctgatacaaa aaaaaaaaaa aaaaaa 599

<210> 13

<211> 115

<212> PRT

<213>-Homo sapiens - .

<400> 13

Cys His Cys Leu Leu His Ser Leu Gln Ser Asp Arg Glu Lys Ser Glu
1 5 10 15

-13-

Gly Leu Pro Val Ala Pro Phe Met Asp Arg Asp Lys Val Thr Lys Ala
 20 25 30

Thr Ala Gln Ile Gly Phe Ile Lys Phe Val Leu Ile Pro Met Phe Glu
 35 40 45

Thr Val Thr Lys Leu Phe Pro Met Val Glu Glu Ile Met Leu Gln Pro
 50 55 60

Leu Trp Glu Ser Arg Asp Arg Tyr Glu Glu Leu Lys Arg Ile Asp Asp
 65 70 75 80

Ala Met Lys Glu Leu Gln Lys Lys Thr Asp Ser Leu Thr Ser Gly Ala
 85 90 95

Thr Glu Lys Ser Arg Glu Arg Ser Arg Asp Val Lys Asn Ser Glu Gly
 100 105 110

Asp Cys Ala
 115

<210> 14

<211> 28

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: primer

<400> 14

agtcgaattc accgtgagaa gtcagaag

28

<210> 15

<211> 28

<212> DNA

<213> Artificial Sequence

- 14 -

<220>

<223> Description of Artificial Sequence: primer

<400> 15

gtcaaagctt acatggtctt gtggtgcc

28

<210> 16

<211> 1303

<212> DNA

<213> Homo sapiens

<220>

<221> CDS

<222> (107) .. (1066)

<400> 16

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cttactaacg ttagcccccga gcctagctat ggagggtgca tgctga gcc ctg gag 115
Ala Leu Glu
1

cac atg tac cac gac ctc ggg ctg gtc agg gac ttc agc atc aac cct 163
 His Met Tyr His Asp Leu Gly Leu Val Arg Asp Phe Ser Ile Asn Pro
 5 10 15

gtc acc ctc agg agg tgg ctg ttc tgc gtc cac gac aac tac aga aac 211
Val Thr Leu Arg Arg Trp Leu Phe Cys Val His Asp Asn Tyr Arg Asn
20 25 30 35

aac ccc ttc cac aac ttc cgg cac tgc ttc tgc gtg gcc cag atg atg 259
 Asn Pro Phe His Asn Phe Arg His Cys Phe Cys Val Ala Gln Met Met
 40 45 50

tac agc atg gtc tgg ctc tgc agt ctc cag gag aag ttc tca caa acg 307
 Tyr Ser Met Val Trp Leu Cys Ser Leu Gln Glu Lys Phe Ser Gln Thr
 55 60 65

gat atc ctg atc cta atg aca gcg gcc atc tgc cac gat ctg gac cat 355
Asp Ile Leu Ile Leu Met Thr Ala Ala Ile Cys His Asp Leu Asp His
70 75 80

ccc ggc tac aac aac acg tac cag atc aat gcc cgc aca gag ctg gcg 403
Pro Gly Tyr Asn Asn Thr Tyr Gln Ile Asn Ala Arg Thr Glu Leu Ala
85 90 95

-15-

gtc cgc tac aat gac atc tca ccg ctg gag aac cac cac tgc gcc gtg	451
Val Arg Tyr Asn Asp Ile Ser Pro Leu Glu Asn His His Cys Ala Val	
100 105 110 115	
 gcc ttc cag atc ctc gcc gag cct gag tgc aac atc ttc tcc aac atc	499
Ala Phe Gln Ile Leu Ala Glu Pro Glu Cys Asn Ile Phe Ser Asn Ile	
120 125 130	
 cca cct gat ggg ttc aag cag atc cga cag gga atg atc aca tta atc	547
Pro Pro Asp Gly Phe Lys Gln Ile Arg Gln Gly Met Ile Thr Leu Ile	
135 140 145	
 ttg gcc act gac atg gca aga cat gca gaa att atg gat tct ttc aaa	595
Leu Ala Thr Asp Met Ala Arg His Ala Glu Ile Met Asp Ser Phe Lys	
150 155 160	
 gag aaa atg gag aat ttt gac tac agc aac gag gag cac atg acc ctg	643
Glu Lys Met Glu Asn Phe Asp Tyr Ser Asn Glu Glu His Met Thr Leu	
165 170 175	
 ctg aag atg att ttg ata aaa tgc tgt gat atc tct aac gag gtc cgt	691
Leu Lys Met Ile Leu Ile Lys Cys Cys Asp Ile Ser Asn Glu Val Arg	
180 185 190 195	
 cca atg gaa gtc gca gag cct tgg gtg gac tgt tta tta gag gaa tat	739
Pro Met Glu Val Ala Glu Pro Trp Val Asp Cys Leu Leu Glu Glu Tyr	
200 205 210	
 ttt atg cag agc gac cgt gag aag tca gaa ggc ctt cct gtg gca ccg	787
Phe Met Gln Ser Asp Arg Glu Lys Ser Glu Gly Leu Pro Val Ala Pro	
215 220 225	
 ttc atg gac cga gac aaa gtg acc aag gcc aca gcc cag att ggg ttc	835
Phe Met Asp Arg Asp Lys Val Thr Lys Ala Thr Ala Gln Ile Gly Phe	
230 235 240	
 atc aag ttt gtc ctg atc cca atg ttt gaa aca gtg acc aag ctc ttc	883
Ile Lys Phe Val Leu Ile Pro Met Phe Glu Thr Val Thr Lys Leu Phe	
245 250 255	
 ccc atg gtt gag gag atc atg ctg cag cca ctt tgg gaa tcc cga gat	931
Pro Met Val Glu Glu Ile Met Leu Gln Pro Leu Trp Glu Ser Arg Asp	
260 265 270 275	
 cgc tac gag gag ctg aag cgg ata gat gac gcc atg aaa gag tta cag	979
Arg Tyr Glu Glu Leu Lys Arg Ile Asp Asp Ala Met Lys Glu Leu Gln	
280 285 290	
 aag aag act gac agc ttg acg tct ggg gcc acc gag aag tcc aga gag	1027
Lys Lys Thr Asp Ser Leu Thr Ser Gly Ala Thr Glu Lys Ser Arg Glu	
295 300 305	

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aga agc aga gat gtg aaa aac agt gaa gga gac tgt gcc tgaggaaagc    1076
Arg Ser Arg Asp Val Lys Asn Ser Glu Gly Asp Cys Ala
          310                      315                      320

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Ser Asn Ile Pro Pro Asp Gly Phe Lys Gln Ile Arg Gln Gly Met Ile
130 135 140

-17-

Thr Leu Ile Leu Ala Thr Asp Met Ala Arg His Ala Glu Ile Met Asp
145 150 155 160

Ser Phe Lys Glu Lys Met Glu Asn Phe Asp Tyr Ser Asn Glu Glu His
165 170 175

Met Thr Leu Leu Lys Met Ile Leu Ile Lys Cys Cys Asp Ile Ser Asn
180 185 190

Glu Val Arg Pro Met Glu Val Ala Glu Pro Trp Val Asp Cys Leu Leu
195 200 205

Glu Glu Tyr Phe Met Gln Ser Asp Arg Glu Lys Ser Glu Gly Leu Pro
210 215 220

Val Ala Pro Phe Met Asp Arg Asp Lys Val Thr Lys Ala Thr Ala Gln
225 230 235 240

Ile Gly Phe Ile Lys Phe Val Leu Ile Pro Met Phe Glu Thr Val Thr
245 250 255

Lys Leu Phe Pro Met Val Glu Glu Ile Met Leu Gln Pro Leu Trp Glu
260 265 270

Ser Arg Asp Arg Tyr Glu Glu Leu Lys Arg Ile Asp Asp Ala Met Lys
275 280 285

Glu Leu Gln Lys Lys Thr Asp Ser Leu Thr Ser Gly Ala Thr Glu Lys
290 295 300

Ser Arg Glu Arg Ser Arg Asp Val Lys Asn Ser Glu Gly Asp Cys Ala
305 310 315 320

<210> 18

<211> 1887

<212> DNA

<213> Homo sapiens

<220>

<221> CDS

<222> (74)..(1672)

<400> 18

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-18-

cgccggggcgc agg atg gga tcc ggc tcc tcc agc tac cgg ccc aag gcc 109
 Met Gly Ser Gly Ser Ser Ser Tyr Arg Pro Lys Ala
 1 5 10

atc tac ctg gac atc gat gga cgc att cag aag gta atc ttc agc aag 157
 Ile Tyr Leu Asp Ile Asp Gly Arg Ile Gln Lys Val Ile Phe Ser Lys
 15 20 25

tac tgc aac tcc agc gac atc atg gac ctg ttc tgc atc gcc acc ggc 205
 Tyr Cys Asn Ser Ser Asp Ile Met Asp Leu Phe Cys Ile Ala Thr Gly
 30 35 40

ctg cct cgg aac acg acc atc tcc ctg ctg acc acc gac gac gcc atg 253
 Leu Pro Arg Asn Thr Thr Ile Ser Leu Leu Thr Thr Asp Asp Ala Met
 45 50 55 60

gtc tcc atc gac ccc acc atg ccc gcg aat tca gaa cgc act ccg tac 301
 Val Ser Ile Asp Pro Thr Met Pro Ala Asn Ser Glu Arg Thr Pro Tyr
 65 70 75

aaa gtg aga cct gtg gcc atc aag caa ctc tcc gag aga gaa gaa tta 349
 Lys Val Arg Pro Val Ala Ile Lys Gln Leu Ser Glu Arg Glu Glu Leu
 80 85 90

atc cag agc gtg ctg gcg cag gtt gca gag cag ttc tca aga gca ttc 397
 Ile Gln Ser Val Leu Ala Gln Val Ala Glu Gln Phe Ser Arg Ala Phe
 95 100 105

aaa atc aat gaa ctg aaa gct gaa gtt gca aat cac ttg gct gtc cta 445
 Lys Ile Asn Glu Leu Lys Ala Glu Val Ala Asn His Leu Ala Val Leu
 110 115 120

gag aaa cgc gtg gaa ttg gaa gga cta aaa gtg gtg gag att gag aaa 493
 Glu Lys Arg Val Glu Glu Gly Leu Lys Val Val Glu Ile Glu Lys
 125 130 135 140

tgc aag agt gac att aag aag atg agg gag gag ctg gcg gcc aga agc 541
 Cys Lys Ser Asp Ile Lys Lys Met Arg Glu Glu Leu Ala Ala Arg Ser
 145 150 155

agc agg acc aac tgc ccc tgt aag tac agt ttt ttg gat aac cac aag 589
 Ser Arg Thr Asn Cys Pro Cys Lys Tyr Ser Phe Leu Asp Asn His Lys
 160 165 170

aag ttg act cct cga cgc gat gtt ccc act tac ccc aag tac ctg ctc 637
 Lys Leu Thr Pro Arg Arg Asp Val Pro Thr Tyr Pro Lys Tyr Leu Leu
 175 180 185

tct cca gag acc atc gag gcc ctg cgg aag ccg acc ttt gac gtc tgg 685
 Ser Pro Glu Thr Ile Glu Ala Leu Arg Lys Pro Thr Phe Asp Val Trp
 190 195 200

ctt tgg gag ccc aat gag atg ctg agc tgc ctg gag cac atg tac cac 733
 Leu Trp Glu Pro Asn Glu Met Leu Ser Cys Leu Glu His Met Tyr His
 205 210 215 220

-19-

gac ctc ggg ctg gtc agg gac ttc agc atc aac cct gtc acc ctc agg	781
Asp Leu Gly Leu Val Arg Asp Phe Ser Ile Asn Pro Val Thr Leu Arg	
225 230 235	
agg tgg ctg ttc tgc gtc cac gac aac tac aga aac aac ccc ttc cac	829
Arg Trp Leu Phe Cys Val His Asp Asn Tyr Arg Asn Asn Pro Phe His	
240 245 250	
aac ttc cgg cac tgc ttc tgc gtg gcc cag atg atg tac agc atg gtc	877
Asn Phe Arg His Cys Phe Cys Val Ala Gln Met Met Tyr Ser Met Val	
255 260 265	
tgg ctc tgc agt ctc cag gag aag ttc tca caa acg gat atc ctg atc	925
Trp Leu Cys Ser Leu Gln Glu Lys Phe Ser Gln Thr Asp Ile Leu Ile	
270 275 280	
cta atg aca gcg gcc atc tgc cac gat ctg gac cat ccc ggc tac aac	973
Leu Met Thr Ala Ala Ile Cys His Asp Leu Asp His Pro Gly Tyr Asn	
285 290 295 300	
aac acg tac cag atc aat gcc cgc aca gag ctg gcg gtc cgc tac aat	1021
Asn Thr Tyr Gln Ile Asn Ala Arg Thr Glu Leu Ala Val Arg Tyr Asn	
305 310 315	
gac atc tca ccg ctg gag aac cac cac tgc gcc gtg gcc ttc cag atc	1069
Asp Ile Ser Pro Leu Glu Asn His His Cys Ala Val Ala Phe Gln Ile	
320 325 330	
ctc gcc gag cct gag tgc aac atc ttc tcc aac atc cca cct gat ggg	1117
Leu Ala Glu Pro Glu Cys Asn Ile Phe Ser Asn Ile Pro Pro Asp Gly	
335 340 345	
ttc aag cag atc cga cag gga atg atc aca tta atc ttg gcc act gac	1165
Phe Lys Gln Ile Arg Gln Gly Met Ile Thr Leu Ile Leu Ala Thr Asp	
350 355 360	
atg gca aga cat gca gaa att atg gat tct ttc aaa gag aaa atg gag	1213
Met Ala Arg His Ala Glu Ile Met Asp Ser Phe Lys Glu Lys Met Glu	
365 370 375 380	
aat ttt gac tac agc aac gag gag cac atg acc ctg ctg aag atg att	1261
Asn Phe Asp Tyr Ser Asn Glu Glu His Met Thr Leu Leu Lys Met Ile	
385 390 395	
ttg ata aaa tgc tgt gat atc tct aac gag gtc cgt cca atg gaa gtc	1309
Leu Ile Lys Cys Cys Asp Ile Ser Asn Glu Val Arg Pro Met Glu Val	
400 405 410	
gca gag cct tgg gtg gac tgt tta tta gag gaa tat ttt atg cag agc	1357
Ala Glu Pro Trp Val Asp Cys Leu Leu Glu Glu Tyr Phe Met Gln Ser	
415 420 425	
gac cgt gag aag tca gaa ggc ctt cct gtg gca ccg ttc atg gac cga	1405
Asp Arg Glu Lys Ser Glu Gly Leu Pro Val Ala Pro Phe Met Asp Arg	
430 435 440	

-20-

gac aaa gtg acc aag gcc aca gcc cag att ggg ttc atc aag ttt gtc 1453
 Asp Lys Val Thr Lys Ala Thr Ala Gln Ile Gly Phe Ile Lys Phe Val
 445 450 455 460

ctg atc cca atg ttt gaa aca gtg acc aag ctc ttc ccc atg gtt gag 1501
 Leu Ile Pro Met Phe Glu Thr Val Thr Lys Leu Phe Pro Met Val Glu
 465 470 475

gag atc atg ctg cag cca ctt tgg gaa tcc cga gat cgc tac gag gag 1549
 Glu Ile Met Leu Gln Pro Leu Trp Glu Ser Arg Asp Arg Tyr Glu Glu
 480 485 490

ctg aag cgg ata gat gac gcc atg aaa gag tta cag aag aag act gac 1597
 Leu Lys Arg Ile Asp Asp Ala Met Lys Glu Leu Gln Lys Lys Thr Asp
 495 500 505

agc ttg acg tct ggg gcc acc gag aag tcc aga gag aga agc aga gat 1645
 Ser Leu Thr Ser Gly Ala Thr Glu Lys Ser Arg Glu Arg Ser Arg Asp
 510 515 520

gtg aaa aac agt gaa gga gac tgt gcc tgaggaaagc ggggggcgtg 1692
 Val Lys Asn Ser Glu Gly Asp Cys Ala
 525 530

gctgcagttc tggacgggct ggccgagctg cgcgggatcc ttgtgcaggg aagagctgcc 1752

ctggggcacct ggcaccacaa gaccatgttt tctaagaacc attttgttca ctgataaaaa 1812

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<212> PRT

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<400> 19

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 35 40 45

-21-

Thr Thr Ile Ser Leu Leu Thr Thr Asp Asp Ala Met Val Ser Ile Asp
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Pro Thr Met Pro Ala Asn Ser Glu Arg Thr Pro Tyr Lys Val Arg Pro
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Val Ala Ile Lys Gln Leu Ser Glu Arg Glu Glu Leu Ile Gln Ser Val
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Leu Ala Gln Val Ala Glu Gln Phe Ser Arg Ala Phe Lys Ile Asn Glu
 100 105 110

Leu Lys Ala Glu Val Ala Asn His Leu Ala Val Leu Glu Lys Arg Val
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Glu Leu Glu Gly Leu Lys Val Val Glu Ile Glu Lys Cys Lys Ser Asp
 130 135 140

Ile Lys Lys Met Arg Glu Glu Leu Ala Ala Arg Ser Ser Arg Thr Asn
 145 150 155 160

Cys Pro Cys Lys Tyr Ser Phe Leu Asp Asn His Lys Lys Leu Thr Pro
 165 170 175

Arg Arg Asp Val Pro Thr Tyr Pro Lys Tyr Leu Leu Ser Pro Glu Thr
 180 185 190

Ile Glu Ala Leu Arg Lys Pro Thr Phe Asp Val Trp Leu Trp Glu Pro
 195 200 205

Asn Glu Met Leu Ser Cys Leu Glu His Met Tyr His Asp Leu Gly Leu
 210 215 220

Val Arg Asp Phe Ser Ile Asn Pro Val Thr Leu Arg Arg Trp Leu Phe
 225 230 235 240

Cys Val His Asp Asn Tyr Arg Asn Asn Pro Phe His Asn Phe Arg His
 245 250 255

Cys Phe Cys Val Ala Gln Met Met Tyr Ser Met Val Trp Leu Cys Ser
 260 265 270

-22-

Leu Gln Glu Lys Phe Ser Gln Thr Asp Ile Leu Ile Leu Met Thr Ala
 275 280 285
 Ala Ile Cys His Asp Leu Asp His Pro Gly Tyr Asn Asn Thr Tyr Gln
 290 295 300
 Ile Asn Ala Arg Thr Glu Leu Ala Val Arg Tyr Asn Asp Ile Ser Pro
 305 310 315 320
 Leu Glu Asn His His Cys Ala Val Ala Phe Gln Ile Leu Ala Glu Pro
 325 330 335
 Glu Cys Asn Ile Phe Ser Asn Ile Pro Pro Asp Gly Phe Lys Gln Ile
 340 345 350
 Arg Gln Gly Met Ile Thr Leu Ile Leu Ala Thr Asp Met Ala Arg His
 355 360 365
 Ala Glu Ile Met Asp Ser Phe Lys Glu Lys Met Glu Asn Phe Asp Tyr
 370 375 380
 Ser Asn Glu Glu His Met Thr Leu Leu Lys Met Ile Leu Ile Lys Cys
 385 390 395 400
 Cys Asp Ile Ser Asn Glu Val Arg Pro Met Glu Val Ala Glu Pro Trp
 405 410 415
 Val Asp Cys Leu Leu Glu Glu Tyr Phe Met Gln Ser Asp Arg Glu Lys
 420 425 430
 Ser Glu Gly Leu Pro Val Ala Pro Phe Met Asp Arg Asp Lys Val Thr
 435 440 445
 Lys Ala Thr Ala Gln Ile Gly Phe Ile Lys Phe Val Leu Ile Pro Met
 450 455 460
 Phe Glu Thr Val Thr Lys Leu Phe Pro Met Val Glu Glu Ile Met Leu
 465 470 475 480
 Gln Pro Leu Trp Glu Ser Arg Asp Arg Tyr Glu Glu Leu Lys Arg Ile
 485 490 495
 Asp Asp Ala Met Lys Glu Leu Gln Lys Lys Thr Asp Ser Leu Thr Ser
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 Glu-Gly Asp-Cys Ala
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<210> 20

<211> 1967

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<212> DNA

<213> Homo sapiens

<220>

<221> CDS

<222> (2) .. (1741)

<400> 20

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tgc aac tcc agc gac atc atg gac ctg ttc tgc atc gcc acc ggc ctg 97
Cys Asn Ser Ser Asp Ile Met Asp Leu Phe Cys Ile Ala Thr Gly Leu
          20             25             30

cct cgg aac acg acc atc tcc ctg ctg acc acc gac gac gcc atg gtc 145
Pro Arg Asn Thr Thr Ile Ser Leu Leu Thr Thr Asp Asp Ala Met Val
          35             40             45

tcc atc gac ccc acc atg ccc gcg aat tca gaa cgc act ccg tac aaa 193
Ser Ile Asp Pro Thr Met Pro Ala Asn Ser Glu Arg Thr Pro Tyr Lys
    50             55             60

gtg aga cct gtg gcc atc aag caa ctc tcc gct gat gtc gag gac aag 241
Val Arg Pro Val Ala Ile Lys Gln Leu Ser Ala Asp Val Glu Asp Lys
    65             70             75             80

aga acc aca agc cgt ggc cag tct gct gag aga cca ctg agg gac aga 289
Arg Thr Thr Ser Arg Gly Gln Ser Ala Glu Arg Pro Leu Arg Asp Arg
          85             90             95

cgg gtt gtg ggc ctg gag cag ccc cgg agg gaa gga gca ttt gaa agt 337
Arg Val Val Gly Leu Glu Gln Pro Arg Arg Glu Gly Ala Phe Glu Ser
    100             105             110

gga cag gta gag ccc agg ccc aga gag ccc cag ggc tgc tac cag gaa 385
Gly Gln Val Glu Pro Arg Pro Arg Glu Pro Gln Gly Cys Tyr Gln Glu
    115             120             125

ggc cag cgc atc cct cca gag aga gaa gaa tta atc cag agc gtg ctg 433
Gly Gln Arg Ile Pro Pro Glu Arg Glu Glu Leu Ile Gln Ser Val Leu
    130             135             140

gcg cag gtt gca gag cag ttc tca aga gca ttc aaa atc aat gaa ctg 481
Ala Gln Val Ala Glu Gln Phe Ser Arg Ala Phe Lys Ile Asn Glu Leu
    145             150             155             160

aaa gct gaa gtt gca aat cac ttg gct gtc cta gag aaa cgc gtg gaa 529
Lys Ala Glu Val Ala Asn His Leu Ala Val Leu Glu Lys Arg Val Glu
          165             170             175

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-24-

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Leu Glu Gly Leu Lys Val Val Glu Ile Glu Lys Cys Lys Ser Asp Ile	
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aag aag atg agg gag gag ctg gcg gcc aga agc agc agg acc aac tgc	625
Lys Lys Met Arg Glu Glu Leu Ala Ala Arg Ser Ser Arg Thr Asn Cys	
195 200 205	
ccc tgt aag tac agt ttt ttg gat aac cac aag aag ttg act cct cga	673
Pro Cys Lys Tyr Ser Phe Leu Asp Asn His Lys Lys Leu Thr Pro Arg	
210 215 220	
cgc gat gtt ccc act tac ccc aag tac ctg ctc tct cca gag acc atc	721
Arg Asp Val Pro Thr Tyr Pro Lys Tyr Leu Leu Ser Pro Glu Thr Ile	
225 230 235 240	
gag gcc ctg cgg aag ccg acc ttt gac gtc tgg ctt tgg gag ccc aat	769
Glu Ala Leu Arg Lys Pro Thr Phe Asp Val Trp Leu Trp Glu Pro Asn	
245 250 255	
gag atg ctg agc tgc ctg gag cac atg tac cac gac ctc ggg ctg gtc	817
Glu Met Leu Ser Cys Leu Glu His Met Tyr His Asp Leu Gly Leu Val	
260 265 270	
agg gac ttc agc atc aac cct gtc acc ctc agg agg tgg ctg ttc tgc	865
Arg Asp Phe Ser Ile Asn Pro Val Thr Leu Arg Arg Trp Leu Phe Cys	
275 280 285	
gtc cac gac aac tac aga aac aac ccc ttc cac aac ttc cgg cac tgc	913
Val His Asp Asn Tyr Arg Asn Asn Pro Phe His Asn Phe Arg His Cys	
290 295 300	
ttc tgc gtg gcc cag atg atg tac agc atg gtc tgg ctc tgc agt ctc	961
Phe Cys Val Ala Gln Met Met Tyr Ser Met Val Trp Leu Cys Ser Leu	
305 310 315 320	
cag gag aag ttc tca caa acg gat atc ctg atc cta atg aca gcg gcc	1009
Gln Glu Lys Phe Ser Gln Thr Asp Ile Leu Ile Leu Met Thr Ala Ala	
325 330 335	
atc tgc cac gat ctg gac cat ccc ggc tac aac aac acg tac cag atc	1057
Ile Cys His Asp Leu Asp His Pro Gly Tyr Asn Asn Thr Tyr Gln Ile	
340 345 350	
aat gcc cgc aca gag ctg gcg gtc cgc tac aat gac atc tca ccg ctg	1105
Asn Ala Arg Thr Glu Leu Ala Val Arg Tyr Asn Asp Ile Ser Pro Leu	
355 360 365	
gag aac cac cac tgc gcc gtg gcc ttc cag atc ctc gcc gag cct gag	1153
Glu Asn His His Cys Ala Val Ala Phe Gln Ile Leu Ala Glu Pro Glu	
370 375 380	
tgc aac atc ttc tcc aac atc cca cct gat ggg ttc aag cag atc cga	1201
Cys Asn Ile Phe Ser Asn Ile Pro Pro Asp Gly Phe Lys Gln Ile Arg	
385 390 395 400	

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cag gga atg atc aca tta atc ttg gcc act gac atg gca aga cat gca 1249
 Gln Gly Met Ile Thr Leu Ile Leu Ala Thr Asp Met Ala Arg His Ala
 405 410 415

gaa att atg gat tct ttc aaa gag aaa atg gag aat ttt gac tac agc 1297
 Glu Ile Met Asp Ser Phe Lys Glu Lys Met Glu Asn Phe Asp Tyr Ser
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aac gag gag cac atg acc ctg ctg aag atg att ttg ata aaa tgc tgt 1345
 Asn Glu Glu His Met Thr Leu Leu Lys Met Ile Leu Ile Lys Cys Cys
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 Asp Ile Ser Asn Glu Val Arg Pro Met Glu Val Ala Glu Pro Trp Val
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gac tgt tta tta gag gaa tat ttt atg cag agc gac cgt gag aag tca 1441
 Asp Cys Leu Leu Glu Glu Tyr Phe Met Gln Ser Asp Arg Glu Lys Ser
 465 470 475 480

gaa ggc ctt cct gtg gca ccg ttc atg gac cga gac aaa gtg acc aag 1489
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 485 490 495

gcc aca gcc cag att ggg ttc atc aag ttt gtc ctg atc cca atg ttt 1537
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gaa aca gtg acc aag ctc ttc ccc atg gtt gag gag atc atg ctg cag 1585
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cca ctt tgg gaa tcc cga gat cgc tac gag gag ctg aag cgg ata gat 1633
 Pro Leu Trp Glu Ser Arg Asp Arg Tyr Glu Glu Leu Lys Arg Ile Asp
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gac gcc atg aaa gag tta cag aag aag act gac agc ttg acg tct ggg 1681
 Asp Ala Met Lys Glu Leu Gln Lys Lys Thr Asp Ser Leu Thr Ser Gly
 545 550 555 560

gcc acc gag aag tcc aga gag aga agc aga gat gtg aaa aac agt gaa 1729
 Ala Thr Glu Lys Ser Arg Glu Arg Ser Arg Asp Val Lys Asn Ser Glu
 565 570 575

gga gac tgt gcc tgaggaaagc ggggggcgtg gctgcagttc tggacgggct 1781
 Gly Asp Cys Ala
 580

ggccgagctg cgccgggatcc-ttgtgcaggg aagagctgcc ctgggcacct_ggcaccacaa_ 1841

gaccatgttt tctaagaacc attttgttca ctgatacaaa aaaaaaaaaa ggaattcatg 1901

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aaaaaa 1967

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<210> 21

<211> 580

<212> PRT

<213> Homo sapiens

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Pro Arg Asn Thr Thr Ile Ser Leu Leu Thr Thr Asp Asp Ala Met Val
      35             40             45

Ser Ile Asp Pro Thr Met Pro Ala Asn Ser Glu Arg Thr Pro Tyr Lys
 50             55             60

Val Arg Pro Val Ala Ile Lys Gln Leu Ser Ala Asp Val Glu Asp Lys
 65             70             75             80

Arg Thr Thr Ser Arg Gly Gln Ser Ala Glu Arg Pro Leu Arg Asp Arg
      85             90             95

Arg Val Val Gly Leu Glu Gln Pro Arg Arg Glu Gly Ala Phe Glu Ser
      100             105             110

Gly Gln Val Glu Pro Arg Pro Arg Glu Pro Gln Gly Cys Tyr Gln Glu
      115             120             125

Gly Gln Arg Ile Pro Pro Glu Arg Glu Glu Leu Ile Gln Ser Val Leu
      130             135             140

Ala Gln Val Ala Glu Gln Phe Ser Arg Ala Phe Lys Ile Asn Glu Leu
      145             150             155             160

Lys Ala Glu Val Ala Asn His Leu Ala Val Leu Glu Lys Arg Val Glu
      165             170             175

Leu Glu Gly Leu Lys Val Val Glu Ile Glu Lys Cys Lys Ser Asp Ile
      180             185             190

Lys Lys Met Arg Glu Glu Leu Ala Ala Arg Ser Ser Arg Thr Asn Cys
      195             200             205

Pro Cys Lys Tyr Ser Phe Leu Asp Asn His Lys Lys Leu Thr Pro Arg
      210             215             220

Arg Asp Val Pro Thr Tyr Pro Lys Tyr Leu Leu Ser Pro Glu Thr Ile
      225             230             235             240

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-27-

Glu Ala Leu Arg Lys Pro Thr Phe Asp Val Trp Leu Trp Glu Pro Asn
 245 250 255
 Glu Met Leu Ser Cys Leu Glu His Met Tyr His Asp Leu Gly Leu Val
 260 265 270
 Arg Asp Phe Ser Ile Asn Pro Val Thr Leu Arg Arg Trp Leu Phe Cys
 275 280 285
 Val His Asp Asn Tyr Arg Asn Asn Pro Phe His Asn Phe Arg His Cys
 290 295 300
 Phe Cys Val Ala Gln Met Met Tyr Ser Met Val Trp Leu Cys Ser Leu
 305 310 315 320
 Gln Glu Lys Phe Ser Gln Thr Asp Ile Leu Ile Leu Met Thr Ala Ala
 325 330 335
 Ile Cys His Asp Leu Asp His Pro Gly Tyr Asn Asn Thr Tyr Gln Ile
 340 345 350
 Asn Ala Arg Thr Glu Leu Ala Val Arg Tyr Asn Asp Ile Ser Pro Leu
 355 360 365
 Glu Asn His His Cys Ala Val Ala Phe Gln Ile Leu Ala Glu Pro Glu
 370 375 380
 Cys Asn Ile Phe Ser Asn Ile Pro Pro Asp Gly Phe Lys Gln Ile Arg
 385 390 395 400
 Gln Gly Met Ile Thr Leu Ile Leu Ala Thr Asp Met Ala Arg His Ala
 405 410 415
 Glu Ile Met Asp Ser Phe Lys Glu Lys Met Glu Asn Phe Asp Tyr Ser
 420 425 430
 Asn Glu Glu His Met Thr Leu Leu Lys Met Ile Leu Ile Lys Cys Cys
 435 440 445
 Asp Ile Ser Asn Glu Val Arg Pro Met Glu Val Ala Glu Pro Trp Val
 450 455 460
 Asp Cys Leu Leu Glu Glu Tyr Phe Met Gln Ser Asp Arg Glu Lys Ser
 465 470 475 480
 Glu Gly Leu Pro Val Ala Pro Phe Met Asp Arg Asp Lys Val Thr Lys
 485 490 495
 Ala Thr Ala Gln Ile Gly Phe Ile Lys Phe Val Leu Ile Pro Met Phe
 500 505 510
 Glu Thr Val Thr Lys Leu Phe Pro Met Val Glu Glu Ile Met Leu Gln
 515 520 525
 Pro Leu Trp Glu Ser Arg Asp Arg Tyr Glu Glu Leu Lys Arg Ile Asp
 530 535 540

-28-

Asp Ala Met Lys Glu Leu Gln Lys Lys Thr Asp Ser Leu Thr Ser Gly
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Ala Thr Glu Lys Ser Arg Glu Arg Ser Arg Asp Val Lys Asn Ser Glu
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Gly Asp Cys Ala
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<210> 22

<211> 1457

<212> DNA

<213> Homo sapiens

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ggaaagtaca gtaaaaagtc cgagtgcagc cgccggggcgc agg atg gga tcc ggc 175
 Met Gly Ser Gly
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tcc tcc agc tac cgg ccc aag gcc atc tac ctg gac atc gat gga cgc 223
 Ser Ser Ser Tyr Arg Pro Lys Ala Ile Tyr Leu Asp Ile Asp Gly Arg
 5 10 15 20

att cag aag gta atc ttc agc aag tac tgc aac tcc agc gac atc atg 271
 Ile Gln Lys Val Ile Phe Ser Lys Tyr Cys Asn Ser Ser Asp Ile Met
 25 30 35

gac ctg ttc tgc atc gcc acc ggc ctg cct cgg aac acg acc atc tcc 319
 Asp Leu Phe Cys Ile Ala Thr Gly Leu Pro Arg Asn Thr Thr Ile Ser
 40 45 50

ctg ctg acc acc gac gac gcc atg gtc tcc atc gac ccc acc atg ccc 367
 Leu Leu Thr Thr Asp Asp Ala Met Val Ser Ile Asp Pro Thr Met Pro
 55 60 65

gcg aat tca gaa cgc act ccg tac aaa gtg aga cct gtg gcc atc aag 415
 Ala Asn Ser Glu Arg Thr Pro Tyr Lys Val Arg Pro Val Ala Ile Lys
 70 75 80

caa ctc tcc gag aga gaa gaa tta atc cag agc gtg ctg gcg cag gtt 463
 Gln Leu Ser Glu Arg Glu Glu Leu Ile Gln Ser Val Leu Ala Gln Val
 85 90 95 100

-29-

gca gag cag ttc tca aga gca ttc aaa atc aat gaa ctg aaa gct gaa	511
Ala Glu Gln Phe Ser Arg Ala Phe Lys Ile Asn Glu Leu Lys Ala Glu	
105 110 115	
ggt gca aat cac ttg gct gtc cta gag aaa cgc gtg gaa ttg gaa gga	559
Val Ala Asn His Leu Ala Val Leu Glu Lys Arg Val Glu Leu Glu Gly	
120 125 130	
cta aaa gtg gtg gag att gag aaa tgc aag agt gac att aag aag atg	607
Leu Lys Val Val Glu Ile Glu Lys Cys Lys Ser Asp Ile Lys Lys Met	
135 140 145	
agg gag gag ctg gcg gcc aga agc agc agg acc aac tgc ccc tgt aag	655
Arg Glu Glu Leu Ala Ala Arg Ser Ser Arg Thr Asn Cys Pro Cys Lys	
150 155 160	
tac agt ttt ttg gat aac cac aag aag ttg act cct cga cgc gat gtt	703
Tyr Ser Phe Leu Asp Asn His Lys Lys Leu Thr Pro Arg Arg Asp Val	
165 170 175 180	
ccc act tac ccc aag tac ctg ctc tct cca gag acc atc gag gcc ctg	751
Pro Thr Tyr Pro Lys Tyr Leu Leu Ser Pro Glu Thr Ile Glu Ala Leu	
185 190 195	
cgg aag ccg acc ttt gac gtc tgg ctt tgg gag ccc aat gag atg ctg	799
Arg Lys Pro Thr Phe Asp Val Trp Leu Trp Glu Pro Asn Glu Met Leu	
200 205 210	
agc tgc ctg gag cac atg tac cac gac ctc ggg ctg gtc agg gac ttc	847
Ser Cys Leu Glu His Met Tyr His Asp Leu Gly Leu Val Arg Asp Phe	
215 220 225	
agc atc aac cct gtc acc ctc agg agg tgg ctg ttc tgc gtc cac gac	895
Ser Ile Asn Pro Val Thr Leu Arg Arg Trp Leu Phe Cys Val His Asp	
230 235 240	
aac tac aga aac aac ccc ttc cac aac ttc cgg cac tgc ttc tgc gtg	943
Asn Tyr Arg Asn Asn Pro Phe His Asn Phe Arg His Cys Phe Cys Val	
245 250 255 260	
gcc cag atg atg tac agc atg gtc tgg ctc tgc agt ctc cag gag aag	991
Ala Gln Met Met Tyr Ser Met Val Trp Leu Cys Ser Leu Gln Glu Lys	
265 270 275	
ttc tca caa acg gat atc ctg atc cta atg aca gcg gcc atc tgc cac	1039
Phe Ser Gln Thr Asp Ile Leu Ile Leu Met Thr Ala Ala Ile Cys His	
280 285 290	
gat ctg gac cat ccc ggc tac aac aac acg tac cag atc aat gcc cgc	1087
Asp Leu Asp His Pro Gly Tyr Asn Asn Thr Tyr Gln Ile Asn Ala Arg	
295 300 305	
aca gag ctg gcg gtc cgc tac aat gac atc tca ccg ctg gag aac cac	1135
Thr Glu Leu Ala Val Arg Tyr Asn Asp Ile Ser Pro Leu Glu Asn His	
310 315 320	

-30-

cac tgc gcc gtg gcc ttc cag atc ctc gcc gag cct gag tgc aac atc 1183
 His Cys Ala Val Ala Phe Gln Ile Leu Ala Glu Pro Glu Cys Asn Ile
 325 330 335 340

 ttc tcc aac atc cca cct gat ggg ttc aag cag atc cga cag gga atg 1231
 Phe Ser Asn Ile Pro Pro Asp Gly Phe Lys Gln Ile Arg Gln Gly Met
 345 350 355

 atc aca tta atc ttg gcc act gac atg gca aga cat gca gaa att atg 1279
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 360 365 370

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 Asp Ser Phe Lys Glu Lys Met Glu Asn Phe Asp Tyr Ser Asn Glu Glu
 375 380 385

 cac atg acc ctg ctg aag atg att ttg ata aaa tgc tgt gat atc tct 1375
 His Met Thr Leu Leu Lys Met Ile Leu Ile Lys Cys Cys Asp Ile Ser
 390 395 400

 aac gag gtc cgt cca atg gaa gtc gca gag cct tgg gtg gac tgt tta 1423
 Asn Glu Val Arg Pro Met Glu Val Ala Glu Pro Trp Val Asp Cys Leu
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<212> PRT

<213> Homo sapiens

<400> 23

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 Ser Asp Ile Met Asp Leu Phe Cys Ile Ala Thr Gly Leu Pro Arg Asn
 35 40 45

 Thr Thr Ile Ser Leu Leu Thr Thr Asp Asp Ala Met Val Ser Ile Asp
 50 55 60

 Pro Thr Met Pro Ala Asn Ser Glu Arg Thr Pro Tyr Lys Val Arg Pro
 65 70 75 80

 Val Ala Ile Lys Gln Leu Ser Glu Arg Glu Glu Leu Ile Gln Ser Val
 85 90 95

-31-

Leu Ala Gln Val Ala Glu Gln Phe Ser Arg Ala Phe Lys Ile Asn Glu
 100 105 110
 Leu Lys Ala Glu Val Ala Asn His Leu Ala Val Leu Glu Lys Arg Val
 115 120 125
 Glu Leu Glu Gly Leu Lys Val Val Glu Ile Glu Lys Cys Lys Ser Asp
 130 135 140
 Ile Lys Lys Met Arg Glu Glu Leu Ala Ala Arg Ser Ser Arg Thr Asn
 145 150 155 160
 Cys Pro Cys Lys Tyr Ser Phe Leu Asp Asn His Lys Lys Leu Thr Pro
 165 170 175
 Arg Arg Asp Val Pro Thr Tyr Pro Lys Tyr Leu Leu Ser Pro Glu Thr
 180 185 190
 Ile Glu Ala Leu Arg Lys Pro Thr Phe Asp Val Trp Leu Trp Glu Pro
 195 200 205
 Asn Glu Met Leu Ser Cys Leu Glu His Met Tyr His Asp Leu Gly Leu
 210 215 220
 Val Arg Asp Phe Ser Ile Asn Pro Val Thr Leu Arg Arg Trp Leu Phe
 225 230 235 240
 Cys Val His Asp Asn Tyr Arg Asn Asn Pro Phe His Asn Phe Arg His
 245 250 255
 Cys Phe Cys Val Ala Gln Met Met Tyr Ser Met Val Trp Leu Cys Ser
 260 265 270
 Leu Gln Glu Lys Phe Ser Gln Thr Asp Ile Leu Ile Leu Met Thr Ala
 275 280 285
 Ala Ile Cys His Asp Leu Asp His Pro Gly Tyr Asn Asn Thr Tyr Gln
 290 295 300
 Ile Asn Ala Arg Thr Glu Leu Ala Val Arg Tyr Asn Asp Ile Ser Pro
 305 310 315 320
 Leu Glu Asn His His Cys Ala Val Ala Phe Gln Ile Leu Ala Glu Pro
 325 330 335
 Glu Cys Asn Ile Phe Ser Asn Ile Pro Pro Asp Gly Phe Lys Gln Ile
 340 345 350
 Arg Gln Gly Met Ile Thr Leu Ile Leu Ala Thr Asp Met Ala Arg His
 355 360 365
 Ala Glu Ile Met Asp Ser Phe Lys Glu Lys Met Glu Asn Phe Asp Tyr
 370 375 380
 Ser Asn Glu Glu His Met Thr Leu Leu Lys Met Ile Leu Ile Lys Cys
 385 390 395 400

-32-

Cys Asp Ile Ser Asn Glu Val Arg Pro Met Glu Val Ala Glu Pro Trp
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Val Asp Cys Leu Leu Glu Glu Tyr Phe Met Gln Ser Asp Arg
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<213> Artificial Sequence

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<223> Description of Artificial Sequence: FLAG epitope

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<211> 18

<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence: primer

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cgaggagtca acttcttg

18

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International Bureau

INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶ : C12N 15/55, 9/16, 5/12, 1/19, 1/21, C07K 16/40, 16/42, G01N 33/566, C12Q 1/68, A61K 38/17	A3	(11) International Publication Number: WO 99/42596 (43) International Publication Date: 26 August 1999 (26.08.99)
(21) International Application Number: PCT/US99/03879 (22) International Filing Date: 23 February 1999 (23.02.99) (30) Priority Data: 60/075,508 23 February 1998 (23.02.98) US (71) Applicant: ICOS CORPORATION [US/US]; 22021 20th Avenue, S.E., Bothell, WA 98021 (US). (72) Inventor: LOUGHNEY, Kate; 7301 34th Avenue, N.E., Seattle, WA 98115 (US). (74) Agent: WILLIAMS, Joseph, A., Jr.; Marshall, O'Toole, Gerstein, Murray & Borun, 6300 Sears Tower, 233 South Wacker Drive, Chicago, IL 60606-6402 (US).		(81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, UZ, VN, YU, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG). Published <i>With international search report.</i> <i>Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i> (88) Date of publication of the international search report: 2 March 2000 (02.03.00)
(54) Title: PHOSPHODIESTERASE 10 (57) Abstract The present invention provides novel human PDE10 polypeptides, polynucleotides encoding the polypeptides, expression constructs comprising the polynucleotides, host cells transformed with the expression constructs; methods for producing PDE10 polypeptides; antisense polynucleotides; and antibodies specifically immunoreactive with the PDE10 polypeptides. The invention further provides methods to identify binding partners of PDE10, and more particularly, binding partners that modulate PDE10 enzyme activity.		

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INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 99/03879

A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 C12N15/55 C12N9/16 C12N5/12 C12N1/19 C12N1/21
C07K16/40 C07K16/42 G01N33/566 C12Q1/68 A61K38/17

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N C07K G01N C12Q A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	EP 0 044 527 A (BYK GULDEN LOMBERG CHEM FAB) 27 January 1982 (1982-01-27) page 5, line 7 - line 10 ---	32, 33
E	WO 99 29873 A (GOODING DOUGLAS H : INCYTE PHARMA INC (US); STREETER DAVID GRAY (US)) 17 June 1999 (1999-06-17) abstract claims 1-21 --- -/--	1-33

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

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"E" earlier document but published on or after the international filing date

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"P" document published prior to the international filing date but later than the priority date claimed

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"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"Z" document member of the same patent family

Date of the actual completion of the international search

10 January 2000

Date of mailing of the international search report

21/01/2000

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2
NL - 2280 HV Rijswijk
Tel: (+31-70) 340-2040. Tx. 31 651 epo nl.
Fax: (+31-70) 340-3016

Authorized officer

Galli, I

INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 99/03879

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No
P,X	<p>FISHER D A ET AL: "Isolation and characterization of PDE9A, a novel human cGMP-specific phosphodiesterase"</p> <p>JOURNAL OF BIOLOGICAL CHEMISTRY,US,AMERICAN SOCIETY OF BIOLOGICAL CHEMISTS, BALTIMORE, MD, vol. 273, no. 25, 19 June 1998 (1998-06-19), page 15559-15564 XP002091363</p> <p>ISSN: 0021-9258</p> <p>the whole document</p> <p style="text-align: center;">---</p>	1-33
P,X	<p>SODERLING S.H. ET AL.: "Identification and characterization of a novel family of cyclic nucleotide phosphodiesterase"</p> <p>J. BIOL. CHEM., vol. 273, no. 25, 19 June 1998 (1998-06-19), pages 15553-15558, XP002127167</p> <p>the whole document</p> <p style="text-align: center;">---</p>	1-33
P,X	<p>DATABASE GENBANK "Online!"</p> <p>Accession AF067225, 18 July 1998 (1998-07-18)</p> <p>GUIPPONI M. ET AL.: "cGMP phosphodiesterase A3"</p> <p>XP002127168</p> <p>the whole document</p> <p style="text-align: center;">---</p>	1-33
A	<p>WO 97 35989 A (INCYTE PHARMA INC ;AU YOUNG JANICE (US); COCKS BENJAMIN GRAEME (US) 2 October 1997 (1997-10-02)</p> <p>abstract</p> <p>claims 1-19</p> <p style="text-align: center;">---</p>	1-33
A	<p>WO 94 28144 A (UNIV WASHINGTON) 8 December 1994 (1994-12-08)</p> <p>abstract</p> <p>page 1-18</p> <p style="text-align: center;">---</p>	1-33
A	<p>WO 92 18541 A (UNIV WASHINGTON) 29 October 1992 (1992-10-29)</p> <p>abstract</p> <p>page 1-38</p> <p style="text-align: center;">-----</p>	1-33

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US 99/03879

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
2. ☒ Claims Nos.:
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
See FURTHER INFORMATION sheet PCT/ISA/210
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims: it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Continuation of Box I.2

Claims 32 and 33 refer to compounds that bind specifically to PDE10 without giving a true technical characterization. Moreover, no such compounds are defined in the application. In consequence, the scope of said claims is ambiguous and vague, and their subject-matter is not sufficiently disclosed and supported (Art. 5 and 6 PCT). No search can be performed for such purely speculative claims whose wording is, in fact, a mere recitation of the results to be achieved.

The applicant's attention is drawn to the fact that claims, or parts of claims, relating to inventions in respect of which no international search report has been established need not be the subject of an international preliminary examination (Rule 66.1(e) PCT). The applicant is advised that the EPO policy when acting as an International Preliminary Examining Authority is normally not to carry out a preliminary examination on matter which has not been searched. This is the case irrespective of whether or not the claims are amended following receipt of the search report or during any Chapter II procedure.

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 99/03879

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
EP 0044527 A	27-01-1982	DE 3027279 A WO 8200293 A EP 0056057 A	06-05-1982 04-02-1982 21-07-1982
WO 9929873 A	17-06-1999	US 5922595 A AU 1710899 A	13-07-1999 28-06-1999
WO 9735989 A	02-10-1997	AU 2542797 A CA 2248675 A EP 0907742 A	17-10-1997 02-10-1997 14-04-1999
WO 9428144 A	08-12-1994	CA 2141060 A EP 0652960 A JP 8502900 T US 5702936 A US 5955583 A US 5652131 A	08-12-1994 17-05-1995 02-04-1996 30-12-1997 21-09-1999 29-07-1997
WO 9218541 A	29-10-1992	CA 2085881 A EP 0535216 A US 5580771 A US 5602019 A US 5800987 A US 5789553 A US 5776752 A US 5389527 A	20-10-1992 07-04-1993 03-12-1996 11-02-1997 01-09-1998 04-08-1998 07-07-1998 14-02-1995